

Defining a research agenda for environmental wastewater surveillance of pathogens

Authors: Alexander G Shaw¹, Catherine Troman¹, Joyce Odeke Akello¹, Kathleen O'Reilly³, Jillian Gaud², Stephanie Grow², Nicholas Grassly¹, Duncan Steele², David Blazes², Supriya Kumar² & the environmental surveillance Working Group*.

Author Affiliations:

¹ Medical Research Council Centre for Global Infectious Disease Analysis, School of Public Health, Imperial College London, London, United Kingdom

² Bill and Melinda Gates Foundation, Seattle, Washington, USA

³ London School of Hygiene & Tropical Medicine

* The Environmental Surveillance Working Group comprises attendees at the *Environmental Surveillance for Public Health Impact Meeting*:

Name	Affiliation
Alex Shaw	Imperial College, London
Catherine Troman	Imperial College, London
Joyce Okello	Imperial College, London
Nicholas Grassly	Imperial College, London
Amrita Sekhar	Bill & Melinda Gates Foundation
Andy Tatem	University of Southampton
Amanda Debes	Johns Hopkins University
Amanda Handley	Murdoch Children's Research Institute
Ana Burgos Gutierrez	HERA
Angela Chaudhuri	Catalyst Management Services
Ben Lepene	Ceres Nanoscience
Ben Pyne	Skoll Foundation
Bradley White	Verily
Christopher Uzzell	Imperial College, London
Cristina M. Tato	Chan Zuckerberg BioHub

Christian Walder	Asian Development Bank
David Larsen	Syracuse University
David Blazes	Bill & Melinda Gates Foundation
Damla Bilgin	Illumina
David Boyle	PATH
Dilip Abraham	Christian Medical College, Vellore
Mahbubur Rahman	Institute of Epidemiology, Disease Control and Research, Bangladesh
Duncan Steele	Bill & Melinda Gates Foundation
Erik Karlsson	Institut Pasteur du Cambodge
Farah Ishtiaq	Tata Institute for Genomes and Society
Farah Qamar	Aga Khan University, Pakistan
Julia Fitzner	WHO, Berlin Hub
Gavin Smith	Duke-NUS, Singapore
Gisela Abbam	Perkin Elmer
Gagandeep Kang	Christian Medical College, Vellore
Hamilton Bennett	Moderna
Hamisu Abdullahi	WHO
Helen Stembridge	UK-Health Services Agency
Imran Nisar	Aga Khan University, Pakistan
Indah Kartika	University of Gadjah Mada
Isobel Blake	Imperial College, London
Ivan Liachko	Phase Genomics
Josie Golding	Wellcome Trust
Julie Bines	Murdoch Children's Research Institute
Jacob John	Christian Medical College, Vellore
Jillian Gauld	Bill & Melinda Gates Foundation
John Dennehy	City University of New York
Joshua Levy	Scripps Research Institute
Jonathan Rigby	Liverpool School of Tropical Medicine
Joshua Trotta	Thermo Fisher
Kathleen O'Reilly	London School of Hygiene and Tropical Medicine
Katrina Kalantar	Chan Zuckerberg BioHub
Kayla Barnes	Liverpool School of Tropical Medicine
Kerrigan McCarthy	National Institute for Communicable Diseases, South Africa
Kim Porter	Bill & Melinda Gates Foundation
Kirsten Vannice	Bill & Melinda Gates Foundation
Karen Menge	Chromacode
Kathie Paul Wilkerson	Skoll Foundation
Kayla Laserson	Bill & Melinda Gates Foundation
Laurette Mhlanga	SACEMA, Stellenbosch University
Lukas von Tobel	Novel-T
Lungi Okoko	Bill & Melinda Gates Foundation

Mariana Matus	BioBot
Marietjie Venter	University of Pretoria
Kate Medlicott	WHO
Michelle Morrison	Bill & Melinda Gates Foundation
Michael Oberholzer	Illumina
Mami Taniuchi	University of Virginia
Mukhlid Yousif	National Institute for Communicable Diseases, South Africa
Nicholas Feasey	Liverpool School of Tropical Medicine
Nicholas Thomson	Sanger Institute
Nitzan Soffer	Illumina
Michael Owusu	KNUST, Ghana
Peter Hart	Wellcome Trust
Raphael Zellweger	International Vaccine Institute, Seoul, Korea
Robbie Barbero	Ceres Nanoscience
Samantha Dolan	Bill & Melinda Gates Foundation
Sampson Twumasi-Ankrah	KNUST, Ghana
Scott Meschke	University of Washington, Seattle
Simon Harris	Bill & Melinda Gates Foundation
Sophie Magnet	PATH
Stephanie Grow	Bill & Melinda Gates Foundation
Stephen Rudd	Oxford Nanopore Technologies
Steve Kroiss	Bill & Melinda Gates Foundation
Supriya Kumar	Bill & Melinda Gates Foundation
Stephane Vouillamoz	Novel-T
Tahmina Shirin	Institute of Epidemiology, Disease Control and Research, Bangladesh
Taslimareif Saiyed	C-CAMP, India
Venkata Raghava Mohan	Christian Medical College, Vellore
Vicka Oktaria	University of Gadjah Mada
Vincent Seaman	Bill & Melinda Gates Foundation
Vanessa Moeder	Illumina
Ben Yaffe	Verily
Yaw Adu-Sarkodie	KNUST, Ghana

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17 Environmental surveillance, defined as the systematic collection of samples and associated infectious
18 disease pathogen data from wastewater for the purpose of informing decisions, has a rich tradition in
19 public health. High-resource settings such as the US¹ and Europe² have started to implement
20 environmental surveillance networks for use of multi-pathogen data from wastewater, including for
21 pandemic preparedness. Implementing environmental surveillance in lower-resource settings, where a
22 large proportion of populations live in houses not connected to convergent sewer systems, has lagged
23 due to epidemiological and resource challenges. Correcting this imbalance is important to ensure
24 equitable access to actionable surveillance.

25 The Bill & Melinda Gates Foundation hosted a meeting in May 2022, bringing together academic,
26 manufacturing, and public health decision-making partners to co-develop a vision for multi-pathogen
27 environmental surveillance, which we report on here. Environmental surveillance could complement
28 clinical surveillance by potentially supporting the detection of multiple pathogens within a single
29 surveillance network at a fraction of the cost of case-based surveillance per capita. During our
30 discussions we focused on seven pathogens that were proposed by the group: poliovirus, *Salmonella*
31 *typhi*, *Vibrio cholerae*, SARS-CoV-2, Hepatitis A and E, and Measles virus. For each of these pathogens,
32 relying on only clinical surveillance to generate actionable data is a challenge for a number of reasons,
33 including a low ratio of symptomatic cases to overall infections (as seen in polio), a lack of gold standard
34 diagnostic technology broadly available (such as for typhoid, cholera, Hepatitis E, and Hepatitis A), and
35 inefficiency of genomic sequencing at the clinic level (which has been the case for SARS-CoV-2 and
36 measles).

37 Optimal sampling sites, frequency, and methods will vary depending on the specific goal of the
38 surveillance program and the pathogen of interest³. Whilst well-mapped sewage networks with
39 enumerated populations can inform sampling site selection in some regions, hydrological maps
40 overlaid with data sets for elevation, bluelines, and population (such as those from WorldPop^{3,4}) are

41 necessary in low-resource settings where open drains or riverine networks receive human waste directly
42 from households. In these areas, site selection approaches are often iterative, due to uncertainties in
43 the data available and connectivity of the networks.

44 Sampling frequency can vary depending on the inference required. Modelling has shown monthly
45 collections to be sufficient for poliovirus detection,⁵ but early warning systems, as desired for SARS-
46 CoV-2 and *Vibrio cholerae*, could require weekly or potentially even more frequent sampling to inform
47 public health action in a timely manner^{6,7}. Sampling frequency may also be driven by: the cost of
48 sampling; travel to and from sites; cold-chain costs; and the capacity of the laboratory to test samples.

49 Choice of sampling method can be driven by the need for sensitivity versus quantitative measurement.

50 Passive or trap sampling (such as using Moore's swabs⁸) can effectively allow greater flow volumes to be
51 sampled but can be difficult to translate to quantitative measurements. Grab samples, by contrast,
52 provide an absolute sampling volume, potentially facilitating quantitative measurements, but sample
53 only at one timepoint. Lowering the cost of automated samplers, optimizing concentration methods,
54 and aligning sampling methods across pathogens are all areas of focus. Thorough testing and
55 standardization will be essential to ensure that the method chosen is suitable for pathogens of interest.

56 The prevalence of target pathogens varies geographically and so nimble, adaptable platforms are
57 optimal for multi-pathogen detection, with customizable TaqMan array cards⁹ and qPCR both
58 demonstrated approaches. However, in some cases genomic sequence data will be required, making a
59 targeted or metagenomic sequencing approach necessary. Direct detection, sequencing, and
60 bioinformatics tools for environmental surveillance that are rapid, adaptable to newly emerging
61 pathogens, cost-effective, and easily deployable also need to be developed.

62 Translating data from environmental surveillance into an assessment that is informative for public
63 health action requires knowledge of the sensitivity and specificity of environmental surveillance, which

64 are affected by the size of the catchment area and sample characteristics (including pH and
65 temperature); but these are not consistently collected. Development of a minimal set of reporting
66 criteria would support inference from environmental surveillance data and drive forward improvements
67 in test accuracy, including limits of detection. Analytical frameworks to integrate information from
68 environmental surveillance and clinical surveillance systems need to be developed. Dashboards
69 developed during the SARS-CoV-2 pandemic are a useful way to rapidly and visually present data and to
70 incorporate environmental surveillance data into national public health systems (as in Dhaka,
71 Bangladesh: <https://dhakaesforsars-cov-2.research.virginia.edu/>), and these public facing tools should
72 be developed further to support epidemiological inference. Action plans should be developed with
73 stakeholders to articulate appropriate actions in response to the combined information from clinical and
74 environmental surveillance data for each situation or setting ¹⁰. Finally, quantification of the costs and
75 benefits of multi-pathogen environmental surveillance are required to support decisions on how these
76 investments should be prioritized. WHO-led review of information provided by environmental
77 surveillance, and guidelines (as has been done for SARS-CoV-2:
78 <https://www.who.int/publications/i/item/WHO-HEP-ECH-WSH-2022.1> , and for polio:
79 <https://apps.who.int/iris/handle/10665/67854>) on operationalizing environmental surveillance and
80 interpreting data from these systems may be useful to aid broader implementation.

81 Sustainable environmental surveillance systems require reliable funding. Identifying funding for multi-
82 pathogen environmental surveillance could be challenging in low-resource settings given constrained
83 government budgets, a potential focus on global health security via a few sentinel systems, and single
84 pathogen-specific funding mechanisms. We are encouraged by the promise of support for multi-
85 pathogen environmental surveillance from the G7 group of nations
86 (<http://www.g7.utoronto.ca/healthmins/2022-0520-communicue.html>). We are also hopeful that
87 networks across low- and high-resource settings where environmental surveillance is being undertaken

88 could lead to shared tools and methodological approaches, ultimately lowering costs at scale for
 89 integrated, multi-pathogen surveillance in low-resource settings.

90 Environmental surveillance has clearly been demonstrated to be useful in polio and SARS-CoV-2, and is
 91 increasingly seen as a viable surveillance system for broader public health use. Methods need to be
 92 optimized across the range of environmental surveillance use-cases and pathogens. In addition, multiple
 93 strands of evidence are required to build the case for integrated, multi-pathogen, environmental
 94 surveillance system (Box 1). Funders, the WHO, nation states, industry partners and academics will need
 95 to coordinate their efforts in order to develop standardized approaches and guidelines for
 96 environmental surveillance, acknowledging the varied contexts of sanitation systems between high- and
 97 low-resource settings.

98 **Box 1. A research and development agenda for environmental surveillance of infectious disease**
 99 **pathogens**

Development of tools for sampling site selection in low-resource, non-sewered networks
Research to better understand how sampling frequency and methods may be aligned across pathogens
Adaptable, cost-effective direct detection, sequencing, and bioinformatics tools for priority pathogens
Minimal criteria for reporting environmental surveillance data
Validation of environmental surveillance for each pathogen by deployment in the field alongside hospital- or clinic-based surveillance that uses gold standard diagnostics
Development of frameworks to integrate environmental surveillance with clinic-based surveillance data
Examine the cost of population-based environmental surveillance, and build the case for sustained funding
Develop best practices for communicating environmental surveillance results to policy makers

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101 **Figure legend**

102 **Figure 1: Use cases may drive measurement goals and environmental surveillance system design.**

103 The goal of poliovirus surveillance is eradication; environmental surveillance, including genomics, is used
 104 to monitor spatial and temporal distributions of both wild type and vaccine-derived viruses, with any
 105 detection leading immediately to vaccination campaigns to prevent disease. SARS-CoV-2 environmental
 106 surveillance has been used to monitor trends and to control outbreaks by informing the use of non-
 107 pharmaceutical interventions; genomics has allowed detection of variants of concern, and the
 108 identification of novel variants. Environmental surveillance of vaccine-preventable diseases such as

109 Typhoid can provide information on transmission levels, and inform vaccine deployment, and allow
110 monitoring of vaccine impact.
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119 Kartika, Isobel Blake, Ivan Liachko, Josie Golding, Julie Bines, Jacob John, Jillian Gauld, John Dennehy,
120 Joshua Levy, Jonathan Rigby, Joshua Trotta, Kathleen O'Reilly, Katrina Kalantar, Kayla Barnes, Kerrigan
121 McCarthy, Kim Porter, Kristen Vannice, Karen Menge, Kathie Paul Wilkerson, Kayla Laserson, Laurette
122 Mhlanga, Lukas von Tobel, Lungi Okoko, Mariana Matus, Marietjie Venter, Megan Diamond, Kate
123 Medicott, Michelle Morrison, Michael Oberholzer, Mami Taniuchi, Mukhlid Yousif, Nick Feasey, Nick
124 Thomson, Nitzan Soffer, Michael Owusu, Peter Hart, Raphael Zellweger, Robbie Barbero, Samantha
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131 Competing interests

132 The authors report no financial or non-financial conflicts of interest.

133 References

- 134 1 Kirby, A. E. *et al.* *MMWR Morb Mortal Wkly Rep* **70**, 1242-1244 (2021).
135 <https://www.ncbi.nlm.nih.gov/pubmed/34499630>
- 136 2 European Commission [https://environment.ec.europa.eu/system/files/2022-
137 10/Proposal%20for%20a%20Directive%20concerning%20urban%20wastewater%20treatment%
138 20%28recast%29.pdf](https://environment.ec.europa.eu/system/files/2022-10/Proposal%20for%20a%20Directive%20concerning%20urban%20wastewater%20treatment%20%28recast%29.pdf) (2022).
- 139 3 Uzzell, C. B. *et al.* Preprint at
140 <https://www.medrxiv.org/content/10.1101/2021.05.21.21257547v1> (2022).
- 141 4 Tatem, A. J. *Epidemics* **40**, 100597 (2022). <https://doi.org:10.1016/j.epidem.2022.100597>
- 142 5 Kalkowska, D. A. *et al.* *Open Forum Infect Dis* **8**, ofab264 (2021).
143 <https://doi.org:10.1093/ofid/ofab264>
- 144 6 Randazzo, W. *et al.* *Water Res* **181**, 115942 (2020).
145 <https://doi.org:10.1016/j.watres.2020.115942>

146 7 Nemudryi, A. *et al.* *Cell Rep Med* **1**, 100098 (2020). <https://doi.org:10.1016/j.xcrm.2020.100098>
147 8 Sikorski, M. J. & Levine, M. M. *Applied and environmental microbiology* **86** (2020).
148 <https://doi.org:10.1128/AEM.00060-20>
149 9 Baker, K. K. *et al.* *Environ Sci Technol* **52**, 10263-10274 (2018).
150 <https://doi.org:10.1021/acs.est.8b01528>
151 10 Public Health England.
152 https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/833211/National_polio_guidelines_2019.pdf (2019).
153
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155