

PERSPECTIVE

Conceptual framework for the definition of preclinical and prodromal frontotemporal dementia

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Abstract

The presymptomatic stages of frontotemporal dementia (FTD) are still poorly defined and encompass a long accrual of progressive biological (preclinical) and then clinical (prodromal) changes, antedating the onset of dementia. The heterogeneity of clinical presentations and the different neuropathological phenotypes have prevented a prior clear description of either preclinical or prodromal FTD. Recent advances in

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therapeutic approaches, at least in monogenic disease, demand a proper definition of these prodementia stages. It has become clear that a consensus lexicon is needed to comprehensively describe the stages that anticipate dementia. The goal of the present work is to review existing literature on the preclinical and prodromal phases of FTD, providing recommendations to address the unmet questions, therefore laying out a strategy for operationalizing and better characterizing these presymptomatic disease stages.

KEYWORDS

definition, frontotemporal dementia, frontotemporal lobar degeneration, mild cognitive and/or behavioral and/or motor impairment, mild cognitive impairment, preclinical, presymptomatic, prodromal

Frontotemporal dementia (FTD) defines a genetically and pathologically heterogeneous group of neurodegenerative disorders with predominant degeneration of the frontal and/or temporal lobes, in which the main neuropathological hallmarks are represented by tau, TAR DNA-binding protein 43 (TDP-43), or fused in sarcoma (FUS) inclusions.^{1,2} Clinically, it is characterized by progressive deterioration in behavior, personality, and/or language, often with parkinsonism and psychiatric features. Different phenotypes have been classically defined on the basis of presenting clinical symptoms: the behavioral variant of FTD (bvFTD), which is associated with early behavioral and personality changes;³ the nonfluent or agrammatic variant of primary progressive aphasia (nfvPPA), with progressive deficits in speech, grammar, and word output; and the semantic variant of PPA (svPPA), a progressive disorder of semantic knowledge and naming.⁴ A significant proportion of patients have associated extrapyramidal symptoms,⁵ which may form part of either a progressive supranuclear palsy (PSP) or corticobasal syndrome (CBS),⁶ and there is considerable clinical overlap with motor neuron disease (MND).⁷

The presymptomatic stages of FTD are still poorly defined and likely encompass a long accrual of progressive biological (preclinical) followed by clinical (prodromal) changes, antedating the onset of dementia. The heterogeneity of clinical presentations and the different neuropathological phenotypes have prevented a prior clear description of either preclinical or prodromal FTD. Recent advances in therapeutic approaches, at least in monogenic disease, make proper definition of these presymptomatic stages more urgent. As postulated for Alzheimer's disease (AD), the ability to intervene early may offer a chance to delay or even prevent neurodegeneration. In AD, the literature has suggested the conceptual framework of a preclinical biologically active process that precedes the onset of a prodromal or mild cognitive impairment (MCI) phase, which is then followed by dementia.^{8,9} The heterogeneous presentation of FTD suggests that a wider set of clinical features might present in the prodromal phase compared to AD. Nonetheless, a similar conceptual framework to MCI could be translated to the FTD field. In this view, we may define a preclinical FTD stage in those subjects with an ongoing neuropathological process but without clinical abnormalities, and a prodromal stage in those subjects with the onset and progression of subtle clinical symptoms.

A privileged point of view for studying the preclinical and prodromal phases of FTD is provided by its genetic forms. Indeed, familial aggregation has been reported in a significant proportion of people with FTD (up to 40% of cases), with mutations in the microtubule-associated protein tau (*MAPT*) and progranulin (*GRN*) gene, or a pathogenic expansion in the chromosome 9 open reading frame 72 (*C9orf72*) as the most common cause of monogenic disease.¹⁰ Mutations in *MAPT* lead to abnormal tau accumulation, while *GRN* and *C9orf72* pathogenic variations are associated with TDP-43 deposition.¹¹ The study of family members bearing a pathogenic mutation has allowed the naturalistic observation of the shift from preclinical and prodromal status to overt disease. There is a wide variation in the age at onset, both within mutation class and within families with the same mutation at least in *GRN* and *C9orf72* mutations,¹² and possible disease modifiers have been recently reported, even though penetrance is high at age 75.¹³

Moreover, several studies have faced the challenge of detecting a clinical, biological, or imaging signature preceding the onset of dementia. A major contribution in this field has been provided by the international consortia devoted to the extensive evaluation of presymptomatic subjects carrying pathogenic mutations. The ongoing European- and Canadian-based Genetic Frontotemporal dementia Initiative (GENFI, www.genfi.org), the US-based Advancing Research & Treatment for Frontotemporal Lobar Degeneration/Longitudinal Evaluation of Familial Frontotemporal Dementia Subjects (ARTFL/LEFFTDS), and the Australian Dominantly Inherited Non-Alzheimer Dementias (DINAD) studies, have recruited cross-sectional and longitudinal data with the aim to identify early alterations in at-risk subjects before the expected onset of disease.^{12,14–16} In addition, the recently established consortia in Latin America (Research Dementia Latin America [ReDLat]) and New Zealand (Genetic FTD Study [FTDGeNZ]) will be able to further contribute to the description of the natural history of the disease.^{17–20} These studies collaborate together under the auspices of the FTD Prevention Initiative (FPI).¹²

It is therefore important for observational studies and clinical trials to determine specific parameters and measures of preclinical and prodromal FTD, to share a common lexicon when identifying patients in the earliest phases of disease. However, several outstanding issues still need proper analysis and scrutiny. To this end, the goal of the

BOX 1 Unmet questions in preclinical and prodromal frontotemporal dementia

1. How do we define the onset of preclinical disease?
2. How do we define further stages of preclinical disease?
3. Is there a "no disease" phase in genetic FTD preceding the onset of preclinical disease?
4. How do we define onset of prodromal disease?
5. How may we assess mild cognitive and/or behavioral and/or motor impairment (MCBMI) due to FTD?
6. How do we include the prodromal neuropsychiatric features (particularly of *C9orf72*) within this framework?
7. How do we include mild features of parkinsonism or motor neuron disease within this scheme?
8. How do we define phenoconversion?
9. What modifies stage and progression of disease?

present work is to review the existing literature on the preclinical and prodromal phases of FTD, discussing and proving recommendations to the nine pressing questions that need a proper definition (see Box 1). This provides a starting point for operationalizing and better characterizing preclinical and prodromal disease stages of FTD. These recommendations should provide guidance for clinical and research applications, particularly at a time when therapeutic clinical trials are focusing on prodromal and preclinical stages of disease, promoting and harmonizing large-scale multicenter collaborative studies, and increasing funding from national and international agencies.

1 | HOW DO WE DEFINE THE ONSET OF PRECLINICAL DISEASE?

The onset of a preclinical disease stage may be theoretically defined by the occurrence of first signs of protein misfolding, presumably initially without either neuronal dysfunction or neurodegeneration, and with no clinical FTD-related symptoms. One of the key questions in the current literature is therefore how we define this switch from a "no disease" stage to a "preclinical stage" with available markers (see Figure 1).

Conceptually, while the disease process may be initiated through misfolded proteins forming neurotoxic oligomers, the first identifiable hallmark of a preclinical disease stage is the abnormal accumulation of pathogenic protein aggregates within cells, including (1) hyperphosphorylated tau, (2) TDP-43 immunoreactive inclusions, (3) FET family proteins (consisting of FUS, Ewing's sarcoma protein [EWS], and TATA-binding protein associated factor 2N [TAF15]), (4) dipeptide repeat proteins (DPR), or (5) still-to-be-defined proteins in those with frontotemporal lobar degeneration-ubiquitin proteasome system (FTLD-UPS) pathology.^{1,21}

Reliable in vivo biomarkers able to predict the two main proteinopathies, namely tau or TDP-43, are not yet available. No TDP-43 positron emission tomography (PET) tracer has been investigated as of yet, and tau PET imaging studies have led to variable results, with the main limitation in the primary tauopathies being the non-specific/off-target binding and variable affinity for different tau species.^{22,23} Sim-

ilarly, fluid biomarkers of tau and TDP-43 in cerebrospinal fluid (CSF) or blood have not shown specificity for FTLD pathology. While blood phosphorylated tau (p-tau₁₈₁ and p-tau₂₁₇) assays have recently been shown to be useful to identify AD, they do not identify primary tauopathies including FTLD.²⁴⁻²⁷ Markers of blood and CSF TDP-43 measurements have been developed but are not specific for TDP-43 pathology.^{28,29} Phosphorylated TDP-43 markers and CSF TDP-43 real-time quaking-induced conversion reaction (RT-QuIC) may improve specificity,^{30,31} but these results await confirmation. TDP-43 aggregates may be found even in a subset of AD patients, or in other neurodegenerative disorders or in some aged people, thus TDP-43 biomarkers may be not completely specific.^{32,33}

Markers for the FET proteins have also not yet been developed. Recent work has identified the presence of a CSF measure that is specific to *C9orf72* expansion carriers. One of the key pathophysiological mechanisms in *C9orf72*-related disease is the accumulation of sense and antisense transcripts of the expanded repeats. These RNA transcripts serve as templates for the synthesis of DPRs through repeat associated non-ATG (RAN) translation. So far, only one of these, the glycine-proline-repeating protein or poly(GP), has been shown to be measurable in CSF,³⁴⁻³⁶ being increased in *C9orf72* expansion carriers in both the presymptomatic and symptomatic phase, and normal in controls. This suggests it could be useful as a preclinical biomarker in genetic FTD.³⁷⁻³⁹ Importantly, reports of autopsy studies in *C9orf72* expansion carriers have also described widespread DPR protein pathology prior to the formation of TDP-43 inclusions and neuronal loss,⁴⁰⁻⁴² suggesting that at least for *C9orf72* expansion carriers, the onset of the preclinical stage is defined by the presence of DPR proteins rather than TDP-43 pathology.

There is also a need for more studies examining the extent of neuropathological findings consistent with FTLD in healthy older people.

Recommendation: The preclinical phase of FTD should theoretically extend from the earliest signs of protein misfolding to the onset of the first clinical symptom of FTD. Based on current knowledge, the onset of a preclinical stage cannot be reliably identified with available biomarkers at this time except potentially for those with *C9orf72* expansions. We recommend that ongoing research aims to identify both PET tracers and fluid biomarkers that can sensitively and specifically show the presence of tau, TDP-43, and FET pathology.

2 | HOW DO WE DEFINE FURTHER STAGES OF PRECLINICAL DISEASE?

The preclinical disease stage may be characterized by when protein accumulation and misfolding is initiated, but later preclinical stages can also be defined. Accumulation of toxic proteins leads to neuronal dysfunction with multiple cellular mechanisms being affected, including the function of mitochondria and stress granules, autophagy, and transcription. The outcome of this is neuronal loss, that is, neurodegeneration. Both dysfunction and loss of neurons occur prior to the onset of clinical symptoms (see Figure 1).

¹⁸F-fluorodeoxyglucose (FDG)-PET detects changes in glucose metabolism in the brain with hypometabolism representing neuronal

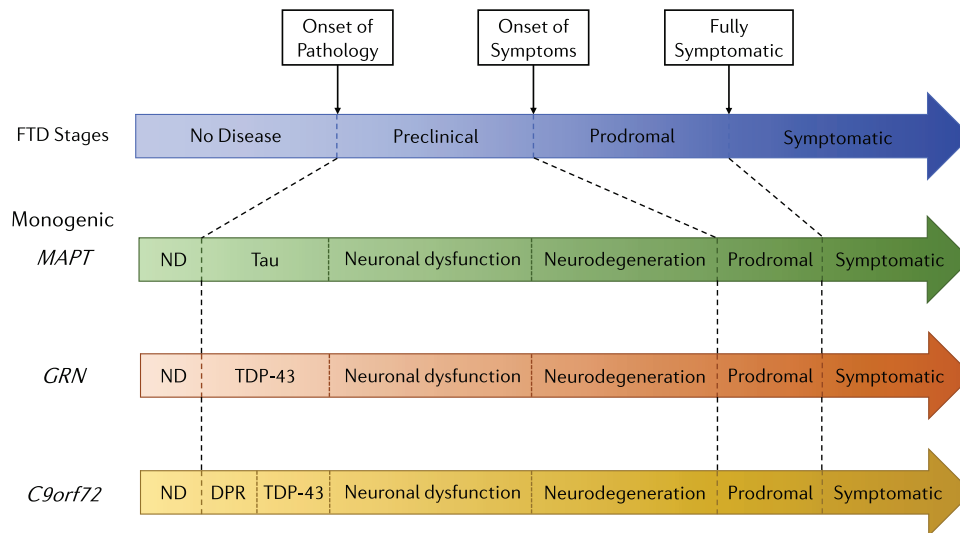


FIGURE 1 Disease stages in frontotemporal dementia (FTD). Natural history of FTD and monogenic FTD subtypes. *C9orf72*, chromosome 9 open reading frame 72; DPR, dipeptide repeat proteins; *GRN*, progranulin; *MAPT*, microtubule-associated protein tau; ND, no disease; TDP-43, TAR DNA-binding protein 43

dysfunction. Studies in AD suggest that FDG-PET may be abnormal prior to neuronal loss measured as atrophy on magnetic resonance imaging (MRI).^{43–46} FDG-PET is also abnormal presymptomatically in genetic FTD,^{47–51} and similar to AD, a few studies have now been performed suggesting that changes occur before structural MRI abnormalities.

Nonetheless, MRI represents one of the most powerful tools to study in vivo neurodegenerative disorders, with a wide range of possible approaches to explore incipient neurodegeneration.^{52,53} The majority of imaging studies in preclinical FTD have used volumetric T1-weighted MRI to investigate changes in gray matter structure and to measure brain volume, the rate of brain atrophy, and the volumes of specific brain regions of interest.^{54–60} In monogenic FTD, volumetric MRI analysis shows significant brain atrophy, first detectable in the insula, at least 10 years before expected symptom onset.¹⁴ Diffusion-weighted MRI detects white matter damage including axonal loss. In genetic FTD, changes to diffusivity have been found in white matter tracts many years before symptom onset.⁶¹ It needs to be further established if and how these subtle changes in gray and white matter found in T1 and diffusion imaging, respectively, may be used as a marker of early neurodegeneration in preclinical stages at the single subject level.

More recent studies have identified a possible fluid biomarker of neurodegeneration, albeit not specific for FTD. Neurofilament light chain (NfL) protein concentrations both in CSF and in blood reflect axonal degeneration and have been shown during the symptomatic period of FTD to be reflective of disease intensity and progression. In the presymptomatic period, analysis seems to suggest that levels change not long prior to symptom onset, increasing by 3- to 4-fold during conversion.^{62–64} While longitudinal NfL measurements could be used to identify mutation carriers approaching symptom onset,⁶⁵ NfL needs to be further studied on a single subject basis, and in particular,

studies showing whether it is sensitive enough to detect neurodegeneration prior to early symptoms (i.e., prior to a prodromal stage).

Recommendation: Neuronal dysfunction can be measured in advance of neuronal loss with FDG-PET imaging but has been poorly studied in presymptomatic FTD thus far. Further studies are important to establish the earliest time at which dysfunction can be detected prior to structural MRI abnormalities, including investigation of newer measures of impaired neuronal function such as novel PET ligands, neurophysiological and magnetoencephalographic markers, and CSF measures of synaptic dysfunction. The onset of neuronal loss may be identifiable by MRI (especially with the advent of ultra-high-field 7T MRI) or fluid biomarkers such as NfL, but it remains unclear which is the most sensitive (early) or specific marker of neurodegeneration in FTD and what cut-offs or thresholds are to be applied, particularly at the single subject level.

3 | IS THERE A “NO DISEASE” PHASE IN GENETIC FTD PRECEDING THE ONSET OF PRECLINICAL DISEASE?

The conceptual timeline of FTD natural history typically includes a healthy stage, with “no disease,” followed by preclinical and prodromal disease to overt dementia (see Figure 1). In monogenic FTD subtypes, some biomarkers appear to be altered from birth and many are abnormal even in young adulthood. This raises the question whether there is a neurodevelopmental dimension to FTD, and the existence of a stage that is without disease, or without neuropathological abnormalities. By analogy with another genetic dementia, Huntington's disease, there may even be fetal neurodevelopmental abnormalities.⁶⁶

Pathogenic loss-of-function mutations in *GRN* lead to haploinsufficiency, with blood and CSF levels of progranulin reduced to < 50% of

normal levels.⁶⁷⁻⁷² Low serum, plasma, or CSF progranulin levels have high accuracy in detecting pathogenic *GRN* mutations,⁷²⁻⁷⁵ with low levels observed from the earliest time period in *GRN* mutation carriers, likely antedating TDP-43 neuropathology. At present, studies have not been performed in children (< 18 years) to understand whether levels are low from birth, but the assumption is that they are, given the known pathophysiology.⁷³⁻⁷⁵

As mentioned above, *C9orf72* expansion carriers have widespread DPR protein pathology early in life.⁴⁰⁻⁴² While similarly to *GRN* mutation carriers studies of fluid biomarkers show abnormal levels (here of raised poly[GP] concentrations) from at least the fourth decade of life,^{37,39,76,77} and no studies have been performed in children, there is a less clear assumption of abnormal levels from birth and studies in a pediatric cohort would be highly informative.

Recommendation: Based on current knowledge it is not clear if a “no disease” stage exists after normal childhood development, for some forms of genetic FTD. For people with *GRN* mutations, there may well be a phase during which a biological disruption is ensuing, but which is not accompanied by an abnormal accumulation of specific pathologic proteins. For people with *C9orf72* expansions, the accumulation of DPRs appears to occur at least in young adulthood, but how early is unknown. Considering also the higher rate of developmental disorders in offspring of patients with FTD,⁷⁸⁻⁸¹ this has suggested the hypothesis of some forms of genetic FTD being neurodevelopmental disorders, in which the boundary with “no disease” is even more indistinct. Studies in pediatric at-risk genetic FTD cohorts, while ethically more complex, will be required to answer these questions more fully.

4 | HOW DO WE DEFINE ONSET OF PRODROMAL DISEASE?

Prodromal FTD may be defined as the presence of subtle cognitive and/or behavioral changes (see Figure 1). Based on studies from large genetic cohorts, the cognitive prodromal phase may start with gradual and progressive executive dysfunction, occurring in isolation or associated with other cognitive changes, such as impaired social cognition or language disturbances. These may be accompanied by behavioral symptoms, such as apathy, disinhibition, loss of empathy, compulsive behavior, and change in appetite or subtle motor deficits,^{14,65,82-90} which are observed by the patient, informant, or clinician, and represent a clear change from the person's usual behavior (see Box 2).

Unlike in AD, for which the concept of MCI was developed to define the prodromal stages,⁹¹⁻⁹³ no detailed characterization of prodromal FTD has been reported. The direct application of the term MCI to FTD is fraught with difficulties given the complex clinical presentation of FTD, which can be heralded by different phenotypes. Attempts to define MCI-like or prodromal stages in FTD have been undertaken with mixed results. Initial criteria for mild behavioral impairment (MBI) excluded serious memory complaints, ignoring cognitive functioning, despite its apparent importance for the early and accurate detection of FTD.^{94,95} The term frontotemporal-MCI (FT-MCI) was later proposed, with criteria including also behavioral symptoms but not requir-

BOX 2 Proposed recommendation for clinical features of prodromal FTD

Gradual and progressive cognitive and/or behavioral and/or motor changes compared to prior functioning and reported by patient or informant, with preservation of independence in functional abilities of daily living, occurring along with one or more of the following features:

- Objective evidence of a dysexecutive syndrome, occurring in isolation or associated with other cognitive changes, such as impaired social cognition, as measured by tests with established specificity for FTD
- Language deficit, as measured by tests with established specificity for FTD
- Behavioral changes: apathy, disinhibition, loss of empathy, compulsive behavior, and change in appetite
- Signs and symptoms of parkinsonism or motor neuron disease

ing the onset to be insidious and progressive, creating potential confusion with delirium, mania, and other conditions.⁹⁶ The phonological similarity in naming with Petersen MCI criteria could also generate confusion.⁹⁷ Finally, provisional MBI criteria have been recently proposed, excluding patients younger than 50 years and not including cognitive disturbances.⁹⁸ Thus, a unifying characterization of prodromal FTD is currently lacking.

Recommendation: The onset of prodromal FTD is characterized by gradual and progressive cognitive and behavioral symptoms, which may be observed by the patient, informant, or clinician, as representing a clear change compared to prior functioning (see Box 2). Given that the onset of prodromal FTD can present with any of behavioral, cognitive, motor or language change, we suggest the label of mild cognitive and/or behavioral and/or motor impairment (MCBMI) to capture the complexity of the clinical phenotype under a single unifying characterization (see next section).

5 | HOW MAY WE ASSESS MCBMI DUE TO FTD?

As with many other neurodegenerative conditions, behavioral and cognitive changes may be present in FTD years before the onset of manifest dementia. These changes clearly describe the switch from preclinical to prodromal disease stage, and a proper description of the first symptoms may further characterize MCBMI due to FTD. Up to now, the most meticulous description of prodromal clinical abnormalities has been performed in at-risk subjects carrying FTD-related pathogenic mutations.⁹⁹

Results from the GENFI study have clearly shown that differences between mutation carriers and non-carriers in neuropsychological measures are apparent about 5 years before the expected onset of dementia, particularly in tests of naming (Boston Naming Test) and executive function (Trail Making Test Part B, Digit Span backward, and Digit Symbol Task), but not in immediate recall and verbal fluency.¹⁴

Previous studies performed in smaller cohorts of presymptomatic mutation carriers obtained somewhat similar findings.^{60,89,100–112}

The wide heterogeneity of clinical presentation and disease progression has so far hindered a clear-cut identification of the core neuropsychological battery tests to adopt for defining MCBMI, both in genetic and in non-monogenic FTD, and for tracking the shift from pre-clinical to prodromal stages. Moreover, for a disease in which behavioral disturbances, including social misconduct, represent the majority of initial symptoms,¹¹³ there is an urgent need to find appropriate standardized tools to detect subtle personality changes preceding the onset of disease.

The assessment of a minimum data set, exportable in different countries, is crucial to define the same outcome measure for clinical trials devoted to delaying or preventing the onset of disease. In this view, a study by the ARTFL/LEFFTDS consortium has shown that the Executive Abilities: Measures and Instruments for Neurobehavioral Evaluation and Research (EXAMINER), a computerized battery developed to quantify many facets of executive functions, is a sensitive measure of cognitive changes in presymptomatic FTD.⁸⁴ Nonetheless, explicit criteria for the use of objective neuropsychological testing are currently lacking and should be defined to harmonize evaluations.

The Clinical Dementia Rating (CDR) plus National Alzheimer's Coordinating Center (NACC) FTLT rating scale (previously called the FTLT-CDR),¹¹⁴ may be a promising measure to identify MCBMI, taking into consideration not only cognitive functions but also language impairment and behavioral and social functioning. Patients with MCI (including mild language impairment) and/or MBI, with relatively preserved functional independence, will have a global score of 0.5. Patients who appear clinically to have a dementia, irrespective of the particular FTD phenotype, will have a global score of ≥ 1 .^{115,116} Recent studies have confirmed the high sensitivity of this scale in identifying patients in the early phases of disease, with very good inter-rater reliability,^{15,84,90,117–120} although the low specificity may limit its use as a screening tool.^{117,118} Nevertheless, the CDR plus NACC FTLT mostly relies on a co-participant/informant report and may lack objectivity. It also does not include measures of neuropsychiatric disturbance.

Recommendation: A provisional definition of MCBMI could rely on the CDR plus NACC FTLT score of 0.5; however, more objective neuropsychological and behavioral measures should be established. Furthermore, any scale aimed at detecting prodromal FTD should incorporate the neuropsychiatric symptoms seen in FTD (see below).

6 | HOW DO WE INCLUDE THE PRODROMAL NEUROPSYCHIATRIC FEATURES (PARTICULARLY OF *C9orf72*) WITHIN THIS FRAMEWORK?

A growing body of evidence describes neuropsychiatric symptoms as early markers of decline along the neurodegenerative spectrum.¹²¹ This is of particular interest in prodromal FTD, in which behavioral symptoms represent the core feature of the disease. What is emerging is that, alongside behavioral symptoms already described in cur-

rent clinical criteria for FTD, such as disinhibition, apathy, loss of empathy, perseverative or compulsive behavior, and hyperorality, other neuropsychiatric symptoms are frequently reported. These manifestations, which are still not defined as FTD core symptoms, should be sought during evaluation and should be considered possible presenting symptoms in the prodromal stages.¹²² In particular, anxiety and depression as well as hallucinations and delusions may be present in people with FTD, the latter highly expressed in *C9orf72* expansion carriers compared to the other FTD subtypes.¹²³ As mentioned above, such features are not captured well by current FTD scales such as the CDR plus NACC FTLT.

More complex, and relevant to the discussion above about the potential neurodevelopmental aspects of *C9orf72*-related FTD, is the presence of apparently lifelong personality traits in people with FTD, including autistic or schizotypy traits, features which may have changed little over time, but must be distinguished from behavioral changes, which evolve and progress over time and that might represent prodromal FTD. The former may end up being scored in symptom rating scales leading to the apparent presence of prodromal symptoms but in reality are not actually changes from a "baseline." These features are important to identify in the earliest FTD stages, allowing a better separation of FTD cases from phenocopies or other mimics.

Recommendation: Further evaluation of the frequency and phenotype of prodromal neuropsychiatric symptoms (particularly in *C9orf72* expansion carriers) is required, with a focus on longstanding autistic and schizotypy traits as well as more overt neuropsychiatric symptoms. Neuropsychiatric evaluation tools will have to consider past psychiatric or personality profiles to reliably identify new emerging prodromal symptoms.

7 | HOW DO WE INCLUDE MILD FEATURES OF PARKINSONISM OR MOTOR NEURON DISEASE WITHIN THIS SCHEME?

A significant percentage of patients with FTD have associated extrapyramidal symptoms, which can be nonspecific, not meeting criteria for a particular disorder, or may fit the criteria for either PSP (Richardson syndrome) or CBS.^{5,6,12,124–131} In both sporadic and genetic FTD, movement disorders can sometimes be the initial presentation.^{132,133} There is also considerable clinical overlap with MND.⁷ Considering that all these diseases are included under the frontotemporal lobar degeneration umbrella term and that most pathogenic mutations may lead to one of these clinical syndromes, initial manifestations of parkinsonism or MND should be identified promptly in the early stages of disease, on par with cognitive and behavioral symptoms. At present, there are no movement disorder scales specific for FTD, although motor behavior may be clinically identified and quantifiable in the prodromal phase by scales designed for other diseases (e.g., the Unified Parkinson's Disease Rating Scale [UPDRS]; Progressive Supranuclear Palsy Rating Scale [PSPRS]; the Amyotrophic Lateral Sclerosis Functional Rating Scale [ALSFRS]).^{65,90,117,118,134}

Recommendation: Motor symptoms are a common feature in FTD, and it may be argued that the onset of isolated movement disorders in the absence of cognitive symptoms could also be defined as a prodromal phase of FTD. We propose a unified approach, potentially including motor features in the prodromal FTD construct, that is, MCBMI. Further studies assessing isolated initial motor symptoms at the onset of sporadic FTD are required.

8 | HOW DO WE DEFINE PHENOCONVERSION?

Applying the definition of “dementia,” namely the presence of cognitive deficits that are significant enough to interfere with instrumental activities of daily living (IADLs), is still challenging in FTD. In early FTD disease stages, patients may present with preservation of IADLs,^{133,135,136} at least as listed for assessment of other disorders, thus not satisfying the diagnosis of dementia, despite the presence of significant behavioral disturbances, executive deficits, or language impairment. Instead of measuring the impact on IADLs, which are somewhat loosely defined in clinical practice and that are useful indicators to track changes from a biological process to a clinical condition in AD, a broader neuropsychiatric approach may be more helpful to define conversion to FTD. In psychiatry, the presence of a mental disorder is defined as a condition that causes significant distress or impairment of personal functioning in social, occupational, or family activities, and must not be merely an expectable response to common stressors and losses.^{137,138}

It is worth noting that the National Institute on Aging–Alzheimer's Association criteria specifically state that a diagnosis of dementia is appropriate in the setting of interference with the ability to function at work or at usual activities, and that the change represents a decline from prior functioning, with changes in personality or behavior plus one other more classic cognitive domains.¹³⁹

As such, conversion to dementia could be defined by symptoms that lead to one or more of the following consequences: (1) the appearance of interference with IADLs, including IADLs relevant to the types of changes induced by FTD; (2) impairment of social/occupational abilities compared to prior functioning, despite preserved autonomy (e.g., normal independence but loss of relationships due to personality changes, inability to hold a job, inadequacy to parent children, language disturbances); (3) a global CDR plus NACC FTLD score ≥ 1 ; (4) fulfillment of consensus criteria for bvFTD or PPA.

The capability to translate abnormal behavior into different social and cultural contexts is yet to be achieved, and transcultural studies defining what is considered socially correct are still lacking. To this end, cooperative and multinational studies are warranted. Furthermore, important implications to consider include extreme behaviors that lead to legal issues such as sexual deviation (paraphilia) or economic difficulties that occur before the detection of a neurodegenerative condition. To this end, co-operative and multinational studies are needed.

Recommendation: The current concept of dementia relies on impairment of IADLs, but this may not be sufficient in defining FTD,

which may comprise impairment of social and occupational functioning adversely impacting a normal lifestyle. Integrating the psychiatric definition of a mental disorder along with the definition of dementia could be an attractive alternative to define the symptomatic phases of FTD and may capture a wider range of conversion.

9 | WHAT MODIFIES STAGE AND PROGRESSION OF DISEASE?

The risk of progression and natural history of preclinical and prodromal FTD may depend on modulating factors, for which the magnitude and interaction have yet to be determined. It has been postulated that certain lifetime experiences, including education, leisure activities, and occupational attainment, may be proxies of cognitive reserve and may modulate brain resistance and resilience.^{140,141} In prodromal FTD, it has been shown that higher educational achievements are associated with greater gray matter volumes, suggesting that subjects with higher education are able to better counteract the detrimental effects of a pathogenetic mutation.¹³ Bilingualism, another emerging aspect of cognitive reserve that has been shown to have an impact also in AD,¹⁴²⁻¹⁴⁴ has been found to delay the onset of dementia in bvFTD but not in PPA.¹⁴⁵ Longitudinal studies have shown that increased education, but also active lifestyles, may also facilitate both brain reserve and brain maintenance in the prodromal stages of genetic FTD,^{146,147} suggesting that cognitive reserve may confer clinical resilience, even in autosomal dominant FTD.

Along with modifiable modulators, even non-modifiable genetic factors have been identified and associated with age at disease onset in FTD. The most established genetic factor, at least in TDP-43 proteinopathies, is the transmembrane protein 106B (*TMEM106B*) gene.¹⁴⁸ It has been suggested that the *TMEM106B* rs1990622 polymorphism might modulate progranulin plasma levels, thus affecting age at symptom onset in *GRN* mutation carriers.^{149,150} Accordingly, subjects with prodromal FTD due to *GRN* mutations and bearing the *TMEM106B* TT genotype showed greater functional brain damage than those with CT/CC *TMEM106B* genotypes.^{13,151} In prodromal FTD-TDP-43 due to *C9orf72* expansion, the relationship is less clear, and it has been suggested that *TMEM106B* might be able to affect disease pathology, but with an opposite association.^{13,152,153} This effect may be an example of the general phenomenon of epistasis, in which a genetic variant is beneficial on some genetic backgrounds but deleterious in others.^{152,154} In the same view, other genetic modifiers, such as apolipoprotein E genotype or *MAPT* haplotypes, should be considered.

Recommendation: Increased cognitive reserve, comprising education, bilingualism, and active lifestyle, are protective factors for FTD progression, in preclinical, prodromal, and dementia phases. The *TMEM106B* TT polymorphism may increase the risk of progression to prodromal FTD in *GRN* carriers. Identification of disease modifiers is key to correctly ranking the risk of disease progression, to stage prodromal FTD and forecast duration of the MCBMI stage, and to select subjects, reducing heterogeneity and increasing statistical power of analysis in clinical trials.¹⁵⁵

10 | CONCLUSIONS AND PERSPECTIVES

Developing the framework of preclinical disease stages as well as MCBMI-FTD continues to pose a challenge, and two aspects should be considered for future studies. On one hand, we should first carefully define the criteria of MCBMI, which may be conceptualized as a “risk state.” MCBMI may represent the prodromal state of FTD, and in some cases, it may refer to a neuropsychiatric condition different from FTD, especially in late-onset cases in which different neuropathologies including AD may coexist, or to a non-progressive or reversible stage. We need a proper definition of clinical features of MCBMI-FTD beyond the label of “mild FTD symptoms”; and to this, reliable biomarkers able to characterize the preclinical and prodromal stages are still clearly needed, as a definition solely based on clinical profile will have low specificity for sporadic cases, particularly in a psychiatric setting. Considering both clinical symptoms and supportive markers, in the near future we may suggest a proper classification of the prodromal stages of FTD to be used in clinical practice and in pharmacological and non-pharmacological trials.

There are some issues that should be considered regarding MCBMI-FTD. FTD is a relatively rare disorder¹⁵⁶ and with a stronger genetic trait than AD.¹² For these reasons, targeting MCBMI-FTD needs further remarks. It is plausible to speculate that markers of preclinical or prodromal FTD in genetic cases at risk of developing disease may be different from what we may observe in overt dementia. A debate is still open on definitions of outcomes in relatively small samples of subjects, with the proposal to identify new personalized endpoints.

The overall considerable proportion of subjects at risk of developing disease due to monogenic mutations, even though still to be established by multinational epidemiological studies,^{156,157} and the possible differences with non-monogenic MCBMI-FTD, raise several questions. First, monogenic disease may help to build up the model of progression from the preclinical to the symptomatic stages. Whether this framework may be applied even in non-monogenic disease, in which the pre-test probability that behavioral or cognitive symptoms will lead to FTD is much lower, needs to be further addressed. Initial findings suggest that clinical presentations (including cognitive, behavioral, and motor) are very similar between genetic and sporadic FTD.^{158,159}

Second, in MCBMI-FTD due to pathogenetic mutations we do not need diagnostic markers, but require prognostic markers, while in sporadic MCBMI-FTD we need both.

Most importantly, we should consider genetic MCBMI-FTD and sporadic MCBMI-FTD as distinct entities regarding treatment approaches. Pathological mutations, that is, *GRN*, *MAPT*, or *C9orf72*, result from specific pathogenetic mechanisms and thus have specific targets of treatment. Conversely, in those cases with unknown pathogenetic mutations, targets for disease-modifying treatments should be centered on the underlying proteinopathy, that is, tau or TDP-43, or nonpharmacological interventions targeting neurotransmitters or connectivity impairment.^{74,160} Conversely, genetic and sporadic MCBMI-FTD can be considered comparable in symptomatic clinical trials and included regardless of the genetic or neuropathological background.

Finally, as with the symptomatic FTD stage, MCBMI-FTD also requires markers of phenotype prediction and markers of proximity to disease onset.

Several issues remain unanswered, including: how do we account for FTD phenocopies; what are the ethical issues in making an earlier diagnosis, informing subjects about biomarkers when it is still uncertain if it will progress to clinical FTD?

All the above considerations represent the roadmap of the recently established GENFI FTD Staging Working Group, whose main objectives will be to answer exhaustively the outstanding issues reported in the present proposal, to identify biomarkers in preclinical and prodromal FTD, and to plan larger collaborative international studies to test the utility and validity of this proposed new approach.

Our ability to carefully characterize the preclinical and prodromal stages of FTD will help in early disease detection, in enabling patient stratification, and in tailoring therapeutic selection for each patient.

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CONFLICTS OF INTEREST

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REFERENCES

1. Neumann M, Mackenzie IRA. Review: neuropathology of non-tau frontotemporal lobar degeneration. *Neuropathol Appl Neurobiol*. 2019;45:19-40.
2. Spillantini MG, Goedert M. Tau pathology and neurodegeneration. *Lancet Neurol*. 2013;12:609-622.
3. Rascovsky K, Hodges JR, Knopman D, et al. Sensitivity of revised diagnostic criteria for the behavioural variant of frontotemporal dementia. *Brain*. 2011;134:2456-2477.
4. Gorno-Tempini ML, Hillis AE, Weintraub S, et al. Classification of primary progressive aphasia and its variants. *Neurology*. 2011;76:1006-1014.
5. Rowe JB. Parkinsonism in frontotemporal dementias. *Int Rev Neurobiol*. 2019;149:249-275.
6. Baizabal-Carvallo JF, Parkinsonism Jankovic J. movement disorders and genetics in frontotemporal dementia. *Nat Rev Neurol*. 2016;12:175-185.
7. Lomen-Hoerth C, Anderson T, Miller B. The overlap of amyotrophic lateral sclerosis and frontotemporal dementia. *Neurology*. 2002;59:1077-1079.
8. Dubois B, Feldman HH, Jacova C, et al. Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria. *Lancet Neurol*. 2014;13:614-629.
9. Jack CR, Bennett DA, Blennow K, et al. NIA-AA Research Framework: toward a biological definition of Alzheimer's disease. *Alzheimers Dement*. 2018;14:535-562.
10. Borroni B, Padovani A. Dementia: a new algorithm for molecular diagnostics in FTL. *Nat Rev Neurol*. 2013;9:241-242.
11. Rademakers R, Neumann M, Mackenzie IR. Advances in understanding the molecular basis of frontotemporal dementia. *Nat Rev Neurol*. 2012;8:423-434.
12. Moore KM, Nicholas J, Grossman M, et al. Age at symptom onset and death and disease duration in genetic frontotemporal dementia: an international retrospective cohort study. *Lancet Neurol*. 2020;19:145-156.
13. Premi E, Grassi M, Van Swieten J, et al. Cognitive reserve and TMEM106B genotype modulate brain damage in presymptomatic frontotemporal dementia: a GENFI study. *Brain*. 2017;140:1784-1791.
14. Rohrer JD, Nicholas JM, Cash DM, et al. Presymptomatic cognitive and neuroanatomical changes in genetic frontotemporal dementia in the Genetic Frontotemporal dementia Initiative (GENFI) study: a cross-sectional analysis. *Lancet Neurol*. 2015;14:253-262.
15. Staffaroni AM, Cobigo Y, Goh S-YM, et al. Individualized atrophy scores predict dementia onset in familial frontotemporal lobar degeneration. *Alzheimer's Dement*. 2020;16:37-48.
16. Pottier C, Zhou X, Perkerson III RB, et al. Potential genetic modifiers of disease risk and age at onset in patients with frontotemporal lobar degeneration and GRN mutations: a genome-wide association study. *Lancet Neurol*. 2018;17:548-558.
17. Ibanez A, Parra MA, Butlerfor C. The Latin America and the Caribbean Consortium on Dementia (LAC-CD): from Networking to Research to Implementation Science. *J Alzheimer's Dis*. 2021: 1-16.
18. Ibanez A, Yokoyama JS, Possin KL, et al. The Multi-Partner Consortium to Expand Dementia Research in Latin America (ReDLat): driving Multicentric Research and Implementation Science. *Front Neurol*. 2021;12:1-16.
19. Parra MA, Baez S, Sedeño L, et al. Dementia in Latin America: paving the way toward a regional action plan. *Alzheimer's Dement*. 2021;17:295-313.
20. Ryan B, Baker A, Ilse C, et al. Diagnosing pre-clinical dementia: the NZ Genetic Frontotemporal Dementia Study (FTDGeNZ). *N Z Med J*. 2018;131:88-91.
21. Mackenzie IR, Neumann M. Subcortical TDP-43 pathology patterns validate cortical FTL-DTP subtypes and demonstrate unique aspects of C9orf72 mutation cases. *Acta Neuropathol*. 2020;139:83-98.
22. Jones DT, Knopman DS, Graff-Radford J, et al. In vivo 18F-AV-1451 tau PET signal in MAPT mutation carriers varies by expected tau isoforms. *Neurology*. 2018;90:e947-54.
23. Bevan-Jones RW, Cope TE, Jones SP, et al. [18 F]AV-1451 binding is increased in frontotemporal dementia due to C9orf72 expansion. *Ann Clin Transl Neurol*. 2018;5:1292-1296.
24. Karikari T, Pascoal T, Ashton N, et al. Plasma phospho-tau181 as a biomarker for Alzheimer's disease: development and validation of a prediction model using data from four prospective cohorts. *Lancet Neurol*. 2020. in press.
25. Janelidze S, Mattsson N, Palmqvist S, et al. Plasma P-tau181 in Alzheimer's disease: relationship to other biomarkers,

- differential diagnosis, neuropathology and longitudinal progression to Alzheimer's dementia. *Nat Med*. 2020;1-8.
26. Thijssen EH, La Joie R, Wolf A, et al. Diagnostic value of plasma phosphorylated tau181 in Alzheimer's disease and frontotemporal lobar degeneration. *Nat Med*. 2020;26:387-397.
 27. Benussi A, Karikari T, Ashton N, et al. Diagnostic and prognostic value of serum NfL and p-Tau181 in frontotemporal lobar degeneration. *J Neurol Neurosurg Psychiatry*. 2020;91:960-967.
 28. Kuiperij HB, Versleijen AAM, Beenes M, et al. Tau rather than TDP-43 proteins are potential cerebrospinal fluid biomarkers for frontotemporal lobar degeneration subtypes: a pilot study. *J Alzheimers Dis*. 2017;55:585-595.
 29. Goossens J, Vanmechelen E, Trojanowski JQ, et al. TDP-43 as a possible biomarker for frontotemporal lobar degeneration: a systematic review of existing antibodies. *Acta Neuropathol Commun*. 2015;3:15.
 30. Suarez-Calvet M, Dols-Icardo O, Lladó A, et al. Plasma phosphorylated TDP-43 levels are elevated in patients with frontotemporal dementia carrying a C9orf72 repeat expansion or a GRN mutation. *J Neurol Neurosurg Psychiatry*. 2014;85:684-691.
 31. Scialò C, Tran TH, Salzano G, et al. TDP-43 real-time quaking induced conversion reaction optimization and detection of seeding activity in CSF of amyotrophic lateral sclerosis and frontotemporal dementia patients. *Brain Commun*. 2020;2,.
 32. Nelson PT, Dickson DW, Trojanowski JQ, et al. Limbic-predominant age-related TDP-43 encephalopathy (LATE): consensus working group report. *Brain*. 2019;142:1503-1527.
 33. Robinson JL, Porta S, Garrett FG, et al. Limbic-predominant age-related TDP-43 encephalopathy differs from frontotemporal lobar degeneration. *Brain*. 2020;143:2844-2857.
 34. Ash PEA, Bieniek KF, Gendron TF, et al. Unconventional translation of C9ORF72 GGGGCC expansion generates insoluble polypeptides specific to c9FTD/ALS. *Neuron*. 2013;77:639-646.
 35. Gendron TF, Bieniek KF, Zhang Y-J, et al. Antisense transcripts of the expanded C9ORF72 hexanucleotide repeat form nuclear RNA foci and undergo repeat-associated non-ATG translation in c9FTD/ALS. *Acta Neuropathol*. 2013;3:829-844.
 36. Mori K, Weng S-M, Arzberger T, et al. The C9orf72 GGGGCC repeat is translated into aggregating dipeptide-repeat proteins in FTL/ALS. *Science*. 2013;339:1335-1338.
 37. Gendron TF, Chew J, Stankowski JN, et al. Poly(GP) proteins are a useful pharmacodynamic marker for C9ORF72-associated amyotrophic lateral sclerosis. *Sci Transl Med*. 2017;9.
 38. Lehmer C, Oeckl P, Weishaupt JH, et al. Poly-GP in cerebrospinal fluid links C9orf72-associated dipeptide repeat expression to the asymptomatic phase of ALS/FTD. *EMBO Mol Med*. 2017;9:859-868.
 39. Meeter LHH, Gendron TF, Sias AC, et al. Poly(GP), neurofilament and grey matter deficits in C9orf72 expansion carriers. *Ann Clin Transl Neurol*. 2018;5:583-597.
 40. Proudfoot M, Gutowski NJ, Edbauer D, et al. Early dipeptide repeat pathology in a frontotemporal dementia kindred with C9ORF72 mutation and intellectual disability. *Acta Neuropathol*. 2014;127:451-458.
 41. Baborie A, Griffiths TD, Jaros E, et al. Accumulation of dipeptide repeat proteins predates that of TDP-43 in frontotemporal lobar degeneration associated with hexanucleotide repeat expansions in C9ORF72 gene. *Neuropathol Appl Neurobiol*. 2015;41:601-612.
 42. Vatsavayai SC, Yoon SJ, Gardner RC, et al. Timing and significance of pathological features in C9orf72 expansion-associated frontotemporal dementia. *Brain*. 2016;139:3202-3216.
 43. Jack CR, Knopman DS, Jagust WJ, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *Lancet Neurol*. 2010;9:119-128.
 44. Jagust W, Gitcho A, Sun F, Kuczynski B, Mungas D, Haan M. Brain imaging evidence of preclinical Alzheimer's disease in normal aging. *Ann Neurol*. 2006;59:673-681.
 45. Reiman EM, Chen K, Alexander GE, et al. Functional brain abnormalities in young adults at genetic risk for late-onset Alzheimer's dementia. *Proc Natl Acad Sci U S A*. 2004;101:284-289.
 46. Kljajevic V, Grothe MJ, Ewers M, Teipel S. Distinct pattern of hypometabolism and atrophy in preclinical and pre-dementia Alzheimer's disease. *Neurobiol Aging*. 2014;35:1973-1981.
 47. De Vocht J, Blommaert J, Devrome M, et al. Use of multimodal imaging and clinical biomarkers in presymptomatic carriers of C9orf72 repeat expansion. *JAMA Neurol*. 2020;77:1008-1017.
 48. Caroppo P, Habert MO, Durrleman S, et al. Lateral temporal lobe: an early imaging marker of the presymptomatic GRN disease?. *J Alzheimer's Dis*. 2015;47:751-759.
 49. Jacova C, Hsiung G-YR, Tawankanjanachot I, et al. Anterior brain glucose hypometabolism predates dementia in progranulin mutation carriers. *Neurology*. 2013;81:1322-1331.
 50. Seelaar H, Papma JM, Garraux G, et al. Brain perfusion patterns in familial frontotemporal lobar degeneration. *Neurology*. 2011;77:384-392.
 51. Deters KD, Risacher SL, Farlow MR, et al. Cerebral hypometabolism and grey matter density in MAPT intron 10 +3 mutation carriers. *Am J Neurodegener Dis*. 2014;3:103-114.
 52. Premi E, Cauda F, Costa T, et al. Looking for neuroimaging markers in frontotemporal lobar degeneration clinical trials: a multi-voxel pattern analysis study in Granulin disease. *J Alzheimers Dis*. 2016;51:249-262.
 53. Feis RA, Bouts MJRJ, Panman JL, et al. Single-subject classification of presymptomatic frontotemporal dementia mutation carriers using multimodal MRI. *Neuroimage Clin*. 2019;22:101718.
 54. Fumagalli GG, Basilico P, Arighi A, et al. Distinct patterns of brain atrophy in Genetic Frontotemporal Dementia Initiative (GENFI) cohort revealed by visual rating scales. *Alzheimers Res Ther*. 2018;10:46.
 55. Bertrand A, Wen J, Rinaldi D, et al. Early cognitive, structural, and microstructural changes in presymptomatic C9orf72 carriers younger than 40 years. *JAMA Neurol*. 2018;75:236-245.
 56. Le Blanc G, Jetté Pomerleau V, McCarthy J, et al. Faster cortical thinning and surface area loss in presymptomatic and symptomatic C9orf72 repeat expansion adult carriers. *Ann Neurol*. 2020;88:113-122.
 57. Filippi M, Agosta F. Mri of non-Alzheimer's dementia: current and emerging knowledge. *Curr Opin Neurol*. 2018;31:405-414.
 58. Panman JL, Jiskoot LC, Bouts MJRJ, et al. Gray and white matter changes in presymptomatic genetic frontotemporal dementia: a longitudinal MRI study. *Neurobiol Aging*. 2019;76:115-124.
 59. Papma JM, Jiskoot LC, Panman JL, et al. Cognition and gray and white matter characteristics of presymptomatic C9orf72 repeat expansion. *Neurology*. 2017;89:1256-1264.
 60. Dopfer EGP, Rombouts SARB, Jiskoot LC, et al. Structural and functional brain connectivity in presymptomatic familial frontotemporal dementia. *Neurology*. 2014;83: e19-26.
 61. Jiskoot LC, Bocchetta M, Nicholas JM, et al. Presymptomatic white matter integrity loss in familial frontotemporal dementia in the GENFI cohort: a cross-sectional diffusion tensor imaging study. *Ann Clin Transl Neurol*. 2018;5:1025-1036.
 62. Meeter LH, Dopfer EG, Jiskoot LC, et al. Neurofilament light chain: a biomarker for genetic frontotemporal dementia. *Ann Clin Transl Neurol*. 2016;3:623-636.
 63. Rostgaard N, Roos P, Portelius E, et al. CSF neurofilament light concentration is increased in presymptomatic CHMP2B mutation carriers. *Neurology*. 2018;90: e157-63.
 64. van der Ende EL, Meeter LH, Poos JM, et al. Serum neurofilament light chain in genetic frontotemporal dementia: a longitudinal, multicentre cohort study. *Lancet Neurol*. 2019;18:1103-1111.

65. Rojas JC, Wang P, Staffaroni AM, et al. Plasma neurofilament light for prediction of disease progression in familial frontotemporal lobar degeneration. *Neurology*. 2021;96:e2296-e2312.
66. Barnat M, Capizzi M, Aparicio E, et al. Huntington's disease alters human neurodevelopment. *Science* (80-). 2020;369:787-793.
67. Van Damme P, Van Hoecke A, Lambrechts D, et al. Progranulin functions as a neurotrophic factor to regulate neurite outgrowth and enhance neuronal survival. *J Cell Biol*. 2008;181:37-41.
68. Baker M, Mackenzie IR, Pickering-Brown SM, et al. Mutations in progranulin cause tau-negative frontotemporal dementia linked to chromosome 17. *Nature*. 2006;442:916-919.
69. Finch N, Baker M, Crook R, et al. Plasma progranulin levels predict progranulin mutation status in frontotemporal dementia patients and asymptomatic family members. *Brain*. 2009;132:583-591.
70. Ghidoni R, Benussi L, Glionna M, Franzoni M, Binetti G. Low plasma progranulin levels predict progranulin mutations in frontotemporal lobar degeneration. *Neurology*. 2008;71:1235-1239.
71. Ghidoni R, Stoppani E, Rossi G, et al. Optimal plasma progranulin cutoff value for predicting null progranulin mutations in neurodegenerative diseases: a multicenter Italian study. *Neurodegener Dis*. 2012;9:121-127.
72. Meeter LHH, Patzke H, Loewen G, et al. Progranulin levels in plasma and cerebrospinal fluid in Granulin mutation carriers. *Dement Geriatr Cogn Dis Extra*. 2016;6:330-340.
73. Galimberti D, Fumagalli GG, Fenoglio C, et al. Progranulin plasma levels predict the presence of GRN mutations in asymptomatic subjects and do not correlate with brain atrophy: results from the GENFI study. *Neurobiol Aging*. 2018;62: 245.e9-245.e12.
74. Benussi A, Gazzina S, Premi E, et al. Clinical and biomarker changes in presymptomatic genetic frontotemporal dementia. *Neurobiol Aging*. 2019;76:133-140.
75. Sellami L, Rucheton B, Ben Younes I, et al. Plasma progranulin levels for frontotemporal dementia in clinical practice: a 10-year French experience. *Neurobiol Aging*. 2020;91:1-9.
76. Cammack AJ, Atassi N, Hyman T, et al. Prospective natural history study of C9orf72 ALS clinical characteristics and biomarkers. *Neurology*. 2019;93: e1605-17.
77. Quaegebeur A, Glaria I, Lashley T, Isaacs AM. Soluble and insoluble dipeptide repeat protein measurements in C9orf72-frontotemporal dementia brains show regional differential solubility and correlation of poly-GR with clinical severity. *Acta Neuropathol Commun*. 2020;8:184.
78. Rogalski E, Johnson N, Weintraub S, Mesulam M. Increased frequency of learning disability in patients with primary progressive aphasia and their first-degree relatives. *Arch Neurol*. 2008;65:244-248.
79. Mesulam MM, Weintraub S. Spectrum of primary progressive aphasia. *Baillieres Clin Neurol*. 1992;1:583-609.
80. Paternicò D, Manes M, Premi E, et al. Frontotemporal dementia and language networks: cortical thickness reduction is driven by dyslexia susceptibility genes. *Sci Rep*. 2016;6: 30848.
81. Alberca R, Montes E, Russell E, Gil-Néciga E, Mesulam M. Left hemispheric hypoplasia in 2 patients with primary progressive aphasia. *Arch Neurol*. 2004;61:265-268.
82. Barker MS, Manoochchri M, Rizer SJ, et al. Recognition memory and divergent cognitive profiles in prodromal genetic frontotemporal dementia. *Cortex*. 2021;139:99-115.
83. Chu SA, Flagan TM, Staffaroni AM, et al. Brain volumetric deficits in MAPT mutation carriers: a multisite study. *Ann Clin Transl Neurol*. 2021;8:95-110.
84. Staffaroni AM, Bajorek L, Casaletto KB, et al. Assessment of executive function declines in presymptomatic and mildly symptomatic familial frontotemporal dementia: nIH-EXAMINER as a potential clinical trial endpoint. *Alzheimer's Dement*. 2020;16:11-21.
85. Malpetti M, Jones PS, Tsvetanov KA, et al. Apathy in presymptomatic genetic frontotemporal dementia predicts cognitive decline and is driven by structural brain changes. *Alzheimer's Dement*. 2020;17:969-983.
86. Tavares TP, Mitchell DGV, Coleman KKL, et al. Early symptoms in symptomatic and preclinical genetic frontotemporal lobar degeneration. *J Neurol Neurosurg Psychiatry*. 2020;91:975-984.
87. Panman JL, Venkatraghavan V, Van Der Ende EL, et al. Modelling the cascade of biomarker changes in GRN -related frontotemporal dementia. *J Neurol Neurosurg Psychiatry*. 2021: 494-501.
88. Benussi A, Premi E, Gazzina S, et al. Progression of behavioral disturbances and neuropsychiatric symptoms in patients with genetic frontotemporal dementia. *JAMA Netw Open*. 2021: 4, 1-17.
89. Jiskoot LC, Panman JL, van Asseldonk L, et al. Longitudinal cognitive biomarkers predicting symptom onset in presymptomatic frontotemporal dementia. *J Neurol*. 2018;265:1381-1392.
90. Olney NT, Ong E, Goh S-YM, et al. Clinical and volumetric changes with increasing functional impairment in familial frontotemporal lobar degeneration. *Alzheimer's Dement*. 2020;16:49-59.
91. Petersen RC, Doody R, Kurz A, et al. Current concepts in mild cognitive impairment. *Arch Neurol*. 2001;58:1985-1992.
92. Petersen RC. Mild Cognitive Impairment. *Continuum (Minneapolis)*. 2016;22:404-418.
93. Albert MS, Dekosky ST, Dickson D, et al. The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *JALZ*. 2011;7:270-279.
94. Taragano FE, Allegri RF, Lyketsos C. Mild behavioral impairment: a prodromal stage of dementia. *Dement Neuropsychol*. 2008;2:256-260.
95. Taragano FE & Allegri R Mild behavioral impairment: the early diagnosis. Presented at the Eleventh International Congress of the International Psychogeriatric Association., Chicago, Illinois: 2003.
96. De Mendonça A, Ribeiro F, Guerreiro M, Garcia C. Frontotemporal mild cognitive impairment. *J Alzheimer's Dis*. 2004;6:1-9.
97. Petersen RC, Smith GE, Waring SC, Ivnik RJ, Tangalos EG, Kokmen E. Mild cognitive impairment. *Arch Neurol*. 1999;56:303.
98. Ismail Z, Smith EE, Geda Y, et al. Neuropsychiatric symptoms as early manifestations of emergent dementia: provisional diagnostic criteria for mild behavioral impairment. *Alzheimer's Dement*. 2016;12:195-202.
99. Greaves CV, Rohrer JD. An update on genetic frontotemporal dementia. *J Neurol*. 2019;266:2075-2086.
100. Geschwind DH, Robidoux J, Alarcón M, et al. Dementia and neurodevelopmental predisposition: cognitive dysfunction in presymptomatic subjects precedes dementia by decades in frontotemporal dementia. *Ann Neurol*. 2001;50:741-746.
101. Janssen JC, Schott JM, Cipolotti L, et al. Mapping the onset and progression of atrophy in familial frontotemporal lobar degeneration. *J Neurol Neurosurg Psychiatry*. 2005;76:162-168.
102. Rohrer JD, Warren JD, Barnes J, et al. Mapping the progression of progranulin-associated frontotemporal lobar degeneration. *Nat Clin Pract Neurol*. 2008;4:455-460.
103. Pievani M, Paternicò D, Benussi L, et al. Pattern of structural and functional brain abnormalities in asymptomatic granulin mutation carriers. *Alzheimer's Dement*. 2014;10:S354-S363.e1.
104. Hallam BJ, Jacova C, Hsiung G-YR, et al. Early neuropsychological characteristics of progranulin mutation carriers. *J Int Neuropsychol Soc*. 2014;20:694-703.
105. Ferman TJ, McRae CA, Arvanitakis Z, Tsuboi Y, Vo A, Wszolek ZK. Early and pre-symptomatic neuropsychological dysfunction in the PPND family with the N279K tau mutation. *Park Relat Disord*. 2003;9:265-270.

106. Spina S, Farlow MR, Unverzagt FW, et al. The tauopathy associated with mutation +3 in intron 10 of Tau: characterization of the MSTD family. *Brain*. 2008;131:72-89.
107. Borroni B, Ghezzi S, Agosti C, et al. Preliminary evidence that VEGF genetic variability confers susceptibility to frontotemporal lobar degeneration. *Rejuvenation Res*. 2008;11:773-780.
108. Borroni B, Alberici A, Cercignani M, et al. Granulin mutation drives brain damage and reorganization from preclinical to symptomatic FTLD. *Neurobiol Aging*. 2012;33:2506-2520.
109. Premi E, Gazzina S, Bozzali M, et al. Cognitive reserve in granulin-related frontotemporal dementia: from preclinical to clinical stages. *PLoS One*. 2013;8: e74762.
110. Premi E, Cauda F, Gasparotti R, et al. Multimodal fMRI resting-state functional connectivity in granulin mutations: the case of frontoparietal dementia. *PLoS One*. 2014;9: e106500.
111. Jiskoot LC, Dopfer EGP, Heijer TD, et al. Presymptomatic cognitive decline in familial frontotemporal dementia: a longitudinal study. *Neurology*. 2016;87:384-391.
112. Montembeault M, Sayah S, Rinaldi D, et al. Cognitive inhibition impairments in presymptomatic C9orf72 carriers. *J Neurol Neurosurg Psychiatry*. 2020;91:366-372.
113. Cheran G, Silverman H, Manoochehri M, et al. Psychiatric symptoms in preclinical behavioural-variant frontotemporal dementia in MAPT mutation carriers. *J Neurol Neurosurg Psychiatry*. 2018;89:449-455.
114. Knopman DS, Kramer JH, Boeve BF, et al. Development of methodology for conducting clinical trials in frontotemporal lobar degeneration. *Brain*. 2008;131:2957-2968.
115. Boeve B, Bove J, Brannelly P, et al. The longitudinal evaluation of familial frontotemporal dementia subjects protocol: framework and methodology. *Alzheimer's Dement*. 2020;16:22-36.
116. Boxer AL, Gold M, Feldman H, et al. New directions in clinical trials for frontotemporal lobar degeneration: methods and outcome measures. *Alzheimer's Dement*. 2020;16:131-143.
117. Miyagawa T, Brushaber D, Syrjanen J, et al. Use of the CDR® plus NACC FTLD in mild FTLD: data from the ARTFL/LEFFTDS consortium. *Alzheimer's Dement*. 2020;16:79-90.
118. Miyagawa T, Brushaber D, Syrjanen J, et al. Utility of the global CDR® plus NACC FTLD rating and development of scoring rules: data from the ARTFL/LEFFTDS Consortium. *Alzheimer's Dement*. 2020;16:106-117.
119. Staffaroni AM, Goh SYM, Cobigo Y, et al. Rates of brain atrophy across disease stages in familial frontotemporal dementia associated with MAPT, GRN, and C9orf72 pathogenic variants. *JAMA Netw Open*. 2020;3: e2022847.
120. Toller G, Ranasinghe K, Cobigo Y, et al. Revised Self-Monitoring Scale: a potential endpoint for frontotemporal dementia clinical trials. *Neurology*. 2020;94: e2384-95.
121. Ismail Z, Agüera-Ortiz L, Brodaty H, et al. The Mild Behavioral Impairment Checklist (MBI-C): a rating scale for neuropsychiatric symptoms in pre-dementia populations. *J Alzheimer's Dis*. 2017;56:929-938.
122. Korhonen T, Katisko K, Cajanus A, et al. Comparison of prodromal symptoms of patients with behavioral variant frontotemporal dementia and Alzheimer disease. *Dement Geriatr Cogn Disord*. 2020;49:98-106.
123. Scarioni M, Gami-Patel P, Timar Y, et al. Frontotemporal dementia: correlations between psychiatric symptoms and pathology. *Ann Neurol*. 2020;87:950-961.
124. Schottlaender LV, Polke JM, Ling H, et al. The analysis of C9orf72 repeat expansions in a large series of clinically and pathologically diagnosed cases with atypical parkinsonism. *Neurobiol Aging*. 2015;36:1221.e1-1221.e6.
125. Gasca-Salas C, Masellis M, Khoo E, et al. Characterization of movement disorder phenomenology in genetically proven, familial frontotemporal lobar degeneration: a systematic review and meta-analysis. *PLoS One*. 2016;11:1-20.
126. Lesage S, Le Ber I, Condroyer C, et al. C9orf72 repeat expansions are a rare genetic cause of parkinsonism. *Brain*. 2013;136:385-391.
127. Choumert A, Poisson A, Honnorat J, et al. G303V tau mutation presenting with progressive supranuclear palsy-like features. *Mov Disord*. 2012;27:581-583.
128. Kelley BJ, Haidar W, Boeve BF, et al. Prominent phenotypic variability associated with mutations in Progranulin. *Neurobiol Aging*. 2009;30:739-751.
129. Snowden JS, Harris J, Adams J, et al. Psychosis associated with expansions in the C9orf72 gene: the influence of a 10 base pair gene deletion. *J Neurol Neurosurg Psychiatry*. 2016;87:562-563.
130. van Swieten JC, Heutink P. Mutations in progranulin (GRN) within the spectrum of clinical and pathological phenotypes of frontotemporal dementia. *Lancet Neurol*. 2008;7:965-974.
131. Le Ber I, Camuzat A, Hannequin D, et al. Phenotype variability in progranulin mutation carriers: a clinical, neuropsychological, imaging and genetic study. *Brain*. 2008;131:732-746.
132. Carneiro F, Saracino D, Huin V, et al. Isolated parkinsonism is an atypical presentation of GRN and C9orf72 gene mutations. *Park Relat Disord*. 2020;80:73-81.
133. Gauthier S, Reisberg B, Zaudig M, et al. Mild cognitive impairment. *Lancet (London, England)*. 2006;367:1262-1270.
134. Stud ALS CNTF. The Amyotrophic Lateral Sclerosis Functional Rating Scale. Assessment of activities of daily living in patients with amyotrophic lateral sclerosis. The ALS CNTF treatment study (ACTS) phase I-II. *Arch Neurol*. 1996;53:141-147.
135. Huey ED, Manly JJ, Tang MX, et al. Course and etiology of dysexecutive MCI in a community sample. *Alzheimer's Dement*. 2013;9:632-639.
136. Kertesz A, McMonagle P, Blair M, Davidson W, Munoz DG. The evolution and pathology of frontotemporal dementia. *Brain*. 2005;128:1996-2005.
137. Stein DJ, Phillips KA, Bolton D, Fulford KWM, Sadler JZ, Kendler KS. What is a mental/psychiatric disorder? from DSM-IV to DSM-V. *Psychol Med*. 2010;40:1759-1765.
138. American Psychiatric Association. *Diagnostic and statistical manual of mental disorders*, 5th ed.: DSM 5. 5th ed. Arlington, VA: 2013.
139. McKhann GM, Knopman DS, Chertkow H, et al. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement*. 2011;7:263-269.
140. Stern Y. Cognitive reserve. *Neuropsychologia*. 2009;47:2015-2028.
141. Stern Y, Barnes CA, Grady C, Jones RN, Raz N. Brain reserve, cognitive reserve, compensation, and maintenance: operationalization, validity, and mechanisms of cognitive resilience. *Neurobiol Aging*. 2019;83:124-129.
142. Perani D, Farsad M, Ballarini T, et al. The impact of bilingualism on brain reserve and metabolic connectivity in Alzheimer's dementia. *Proc Natl Acad Sci U S A*. 2017;114:1690-1695.
143. Mendez MF. Bilingualism and dementia: cognitive reserve to linguistic competency. *J Alzheimer's Dis*. 2019;71:377-388.
144. Ramakrishnan S, Mekala S, Mamidipudi A, et al. Comparative effects of education and bilingualism on the onset of mild cognitive impairment. *Dement Geriatr Cogn Disord*. 2017;44:222-231.
145. Alladi S, Bak TH, Shailaja M, et al. Bilingualism delays the onset of behavioral but not aphasic forms of frontotemporal dementia. *Neuropsychologia*. 2017;99:207-212.
146. Gazzina S, Grassi M, Premi E, et al. Education modulates brain maintenance in presymptomatic frontotemporal dementia. *J Neurol Neurosurg Psychiatry*. 2019;1-7.

147. Casaletto KB, Staffaroni AM, Wolf A, et al. Active lifestyles moderate clinical outcomes in autosomal dominant frontotemporal degeneration. *Alzheimer's Dement.* 2020;16:91-105.
 148. Van Deerlin VM, Sleiman PMA, Martinez-Lage M, et al. Common variants at 7p21 are associated with frontotemporal lobar degeneration with TDP-43 inclusions. *Nat Genet.* 2010;42:234-239.
 149. Finch N, Benussi L, Carrasquillo MM, et al. TMEM106B regulates progranulin levels and the penetrance of FTL in GRN mutation carriers. *Neurology.* 2011;76:467-474.
 150. Cruchaga C, Graff C, Chiang H-H, et al. Association of TMEM106B gene polymorphism with age at onset in granulin mutation carriers and plasma granulin protein levels. *Arch Neurol.* 2011;68:581-586.
 151. Premi E, Formenti A, Gazzina S, et al. Effect of TMEM106B polymorphism on functional network connectivity in asymptomatic GRN mutation carriers. *JAMA Neurol.* 2014;71:216-221.
 152. Gallagher MD, Suh E, Grossman M, et al. TMEM106B is a genetic modifier of frontotemporal lobar degeneration with C9orf72 hexanucleotide repeat expansions. *Acta Neuropathol.* 2014;127:407-418.
 153. van Blitterswijk M, Mullen B, Nicholson AM, et al. TMEM106B protects C9ORF72 expansion carriers against frontotemporal dementia. *Acta Neuropathol.* 2014;127:397-406.
 154. Busch JI, Unger TL, Jain N, Skrinak RT, Charan RA, Chen-Plotkin AS. Increased expression of the frontotemporal dementia risk factor TMEM106B causes C9orf72-dependent alterations in lysosomes. *Hum Mol Genet.* 2016;25:2681-2697.
 155. Wauters E, Van Mossevelde S, Van der Zee J, Cruts M, Van Broeckhoven C. Modifiers of GRN-associated frontotemporal lobar degeneration. *Trends Mol Med.* 2017;23:962-979.
 156. Logroscino G, Piccininni M, Binetti G, et al. Incidence of frontotemporal lobar degeneration in Italy: the Salento-Brescia Registry study. *Neurology.* 2019;92:e2355-63.
 157. Coyle-Gilchrist ITS, Dick KM, Patterson K, et al. Prevalence, characteristics, and survival of frontotemporal lobar degeneration syndromes. *Neurology.* 2016;86:1736-1743.
 158. Heuer HW, Wang P, Rascovsky K, et al. Comparison of sporadic and familial behavioral variant frontotemporal dementia (FTD) in a North American cohort. *Alzheimer's Dement.* 2020;16:60-70.
 159. Capozzo R, Sassi C, Hammer MB, et al. Clinical and genetic analyses of familial and sporadic frontotemporal dementia patients in Southern Italy. *Alzheimer's Dement.* 2017;13:858-869.
 160. Benussi A, Grassi M, Palluzzi F, et al. Classification accuracy of transcranial magnetic stimulation for the diagnosis of neurodegenerative dementias. *Ann Neurol.* 2020;87:394-404.
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APPENDIX A

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