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# Assessing for Cardiotoxicity from Metal-on-Metal Hip Implants with Advanced Multimodality Imaging Techniques

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### Background:

High failure rates of metal-on-metal (MoM) hip implants prompted regulatory authorities to issue worldwide safety alerts. Circulating cobalt from these implants causes rare but fatal autopsy-diagnosed cardiotoxicity. There is concern that milder cardiotoxicity may be common and underrecognized. Although blood metal ion levels are easily measured and can be used to track local toxicity, there are no noninvasive tests for organ deposition. We sought to detect and **constrain** correlation between blood metal ions and a comprehensive panel of established markers of early cardiotoxicity.

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### Methods:

Ninety patients were recruited into this prospective single-center blinded study. Patients were divided into 3 age and sex-matched groups according to implant type and whole-blood metal ion levels. Group-A patients had a ceramic-on-ceramic [CoC] bearing; Group B, an MoM bearing and low blood metal ion levels; and Group C, an MoM bearing and high blood metal-ion levels. All patients underwent detailed cardiovascular phenotyping using cardiac magnetic resonance imaging (CMR) with T2\*, T1, and extracellular volume mapping; echocardiography; and cardiac blood biomarker sampling. T2\* is a novel CMR biomarker of tissue metal loading. Primary outcomes were prespecified on trial registration (ClinicalTrials.gov: NCT02331264).

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### Results:

Blood cobalt levels differed significantly among groups A, B, and C (mean and standard deviation [SD],  $0.17 \pm 0.08$ ,  $2.47 \pm 1.81$ , and  $30.0 \pm 29.1$  ppb, respectively) and between group A and groups B and C combined. No significant between-group differences were found in the left atrial or ventricle size, ejection fraction (on CMR or echocardiography), T1 or T2\* values, extracellular volume, B-type natriuretic peptide level, or troponin level, and all values were within normal ranges. There was no relationship between cobalt levels and ejection fraction ( $R = 0.022$ , 95% confidence interval [CI] =  $-0.185$  to  $0.229$ ) or T2\* values ( $R = -0.108$ , 95% CI =  $-0.105$  to  $0.312$ ).

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### Conclusions:

Using the best available technologies, we did not find that high (but not extreme) blood cobalt and chromium levels had any significant cardiotoxic effect on patients with an MoM hip implant. There were negligible-to-weak correlations between elevated blood metal ion levels and ejection fraction even at the extremes of the 95% CI, which excludes any clinically important association.

### Level of Evidence:

Therapeutic Level II. See Instructions for Authors for a complete description of levels of evidence.

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Over one million patients worldwide have implanted metal-on-metal (MoM) hip prostheses, predominantly composed of cobalt-chromium alloy<sup>1</sup>. Release of nano-particulate debris and metal ions from implants has been reported to cause harm<sup>2</sup>. Local soft-tissue deposition adjacent to the joint<sup>3-5</sup> results in high failure rates of MoM hip implants. This led regulatory agencies to issue safety alerts against their use<sup>6-8</sup>, necessitating surveillance that includes serial measurement of blood metal ion levels and cross-sectional imaging to guide revision surgery<sup>9</sup>.

The relationship between MoM hip implants and systemic toxicity is less clear. Elevated levels of circulating cobalt have been linked with cardiac, thyroid, and neuro-ocular abnormalities in case reports, but causation and prevalence remain to be established<sup>10-18</sup>. Diagnosis of cardiotoxicity requires invasive myocardial biopsy or postmortem examination, with no available noninvasive methods for measuring metal deposition. Heart failure is common in the patient population most likely to undergo hip replacement<sup>19</sup>, so there is concern that an association between MoM hip prostheses and heart failure may be missed. One echocardiography study showed a 7% reduction in left ventricular ejection fraction in MoM-implant recipients<sup>20</sup> (although all values were within the normal range). More recently, an epidemiological study that compared 121 subjects who had 1 type of MoM hip prosthesis with 3,546 subjects who had a metal-on-plastic (MoP) prosthesis showed the age-adjusted rate of hospitalization for heart failure to be equivalent to 1 such hospitalization for every 11 patients<sup>21</sup>. The Therapeutic Goods Administration (Australian Department of Health) consequently published a safety alert pending further investigation<sup>22</sup>.

Cardiac magnetic resonance imaging (CMR) is the gold-standard technique for measuring cardiac volumes and function. In addition, the CMR T2\* method for assessing cardiac iron has been histologically validated and has become a routine measure, transforming the management of patients at risk for iron cardiomyopathy<sup>23,24</sup>. As cobalt exhibits a magnetic property similar to that of iron, T2\* mapping has the potential to noninvasively detect tissue deposition of cobalt in patients with a failing MoM hip prosthesis, as recently demonstrated by our group<sup>25</sup>.

The present study of patients with an MoM hip prosthesis was designed to assess the effect of elevated whole-blood metal ion levels on cardiac function as well as to seek evidence of cardiac metal ion deposition with gold-standard multimodality imaging and measurement of serum biomarkers to minimize potential bias.

## Materials and Methods

### Study Design

In this single-center blinded cross-sectional study, all patients underwent CMR, transthoracic echocardiography, and blood sampling during a single visit (Fig. 1). The research was approved by the institutional review board and ethics committee (reference: 14/LO/1722) and was registered in ClinicalTrials.gov (NCT02331264).

### Setting

Between October 2014 and November 2015, 108 patients were recruited from specialist outpatient clinics held at the Royal National Orthopaedic Hospital and cardiac assessment was performed at the Heart Hospital (both University College London hospitals, U.K.).

### Patient Groups

Patients older than 18 years of age with an MoM or ceramic-on-ceramic (CoC) hip implant in situ for >12 months were recruited. They were divided into 3 age and sex-matched groups on the basis of prosthesis type and blood metal ion levels. Group A consisted of

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patients with a CoC-bearing implant and normal whole-blood metal ion levels; Group B, patients with an MoM implant and low whole-blood metal ion levels (<7 ppb); and Group C, patients with an MoM implant and raised whole-blood metal ion levels ( $\geq 7$  ppb). CoC was chosen over MoP couplings in Group A because of concerns regarding metal debris arising through trunnionosis in MoP implants, and 7 ppb was used as a cutoff point between Groups B and C as this represents the U.K. Medicines & Healthcare products Regulatory Agency (MHRA) recommended threshold above which there may be heightened concern<sup>6</sup>.

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Exclusion criteria were prior hip revision surgery, known atrial fibrillation or impaired renal function (estimated glomerular filtration rate of <30 mL/min), and standard contraindications to magnetic resonance imaging (MRI) (e.g., a pacemaker). Six patients—4 of whom would have been assigned to group A (CoC) and 1 each of whom, to groups B and C—were excluded because of arrhythmias (e.g., atrial fibrillation).

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All participants gave written informed consent conforming to the Declaration of Helsinki (5th revision, 2000). All tests were performed at a single visit. Study data were collected and managed using REDCap (Research Electronic Data Capture software, version 5.9.6, <http://www.project-redcap.org/>), with the cardiac investigators blinded to the study groups during data acquisition and analysis until study completion.

### ***Transthoracic Echocardiography***

Echocardiography was performed using a Vivid E9 ultrasound machine (GE Healthcare). Measurements were made according to the British Society of Echocardiography standard protocol<sup>26</sup>, including acquisition of standard 2-dimensional views, M-mode, spectral tissue Doppler imaging, and blood flow measurements (Fig. 2-A).

### ***CMR Acquisition***

Patients underwent CMR at 1.5 T (Avanto; Siemens Medical). Cardiac volumes and ejection fractions were calculated conventionally from short-axis cine images.

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Bright-blood T2\* and precontrast T1 mapping of the myocardium and liver were acquired on a mid-left ventricular short axis slice and an axial mid-hepatic slice, respectively (Figs. 2-C and 2-D). As T1 mapping is not fully standardized, we used 2 implementations: modified look-locker (MOLLI) and shortened MOLLI (ShMOLLI) sequences (MyoMaps; Siemens). Late gadolinium-enhancement images were acquired using a motion-corrected phase sensitive inversion recovery sequence to identify focal myocardial fibrosis after administering 0.1 mmol/kg of gadolinium-based contrast medium (gadoterate meglumine [Dotarem; Guerbet]). Fifteen minutes after the injection of the contrast medium, T1 sequences were repeated for extracellular volume quantification (Figs. 3-A, 3-B, and 3-C).

### ***Blood Biomarkers***

Blood sampling to measure B-type natriuretic peptide (BNP), troponin-I, and whole-blood cobalt and chromium levels (Fig. 2-B) was performed before the patients underwent CMR. BNP and high-sensitivity troponin are the most commonly used biomarkers in cardiology. They cover a range of cardiac processes, with BNP measurements used to evaluate myocyte strain and troponin measurements indicating myocyte death. Both have prognostic importance.

Whole-blood cobalt and chromium levels were measured using inductively coupled plasma mass spectrometry in the same U.K. reference laboratory to eliminate interlaboratory variation.

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### ***Image Analysis***

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All image acquisition and analyses were performed by observers blinded to the study groups. Unblinding was performed by an independent statistician once all data had been acquired and analyzed, with the data set locked.

Left ventricular volumes, ejection fraction, and mass were calculated from CMR data using standard techniques and dedicated software (CMRtools; Cardiovascular Imaging Solutions). Thresholding methods were used, and papillary muscles were considered part of the left ventricular myocardium. Volumes were subsequently indexed to body surface area. Two of the authors determined the presence of late gadolinium enhancement through visual assessment.

For T2\* and T1 measurements, a region of interest was manually drawn on the interventricular septum on each image, with care taken to avoid the endocardial and epicardial contours to minimize partial voluming effect. Extracellular volume was calculated using MOLLI sequences via a fully automated method that calculates pixel-wise extracellular volume parametric maps<sup>27</sup>, based on the standard formula:

$$\text{extracellular volume} = (1 - \text{hematocrit}) \times (\Delta R1_{\text{myocardium}} / \Delta R1_{\text{blood}})$$

where  $\Delta R1$  is the change in R1 from the postcontrast to the precontrast image, and  $R1 = 1/T1$ .

Echocardiographic data were digitally stored for offline analysis with EchoPAC dimension software (GE Healthcare). Conventional analysis of left ventricular structure and systolic and diastolic function including spectral tissue Doppler parameters were performed according to guidelines, with values averaged over 2 cardiac cycles. Tissue Doppler parameters for lateral and septal walls were averaged to produce a single marker of systolic longitudinal function.

### Study Size

Sample size calculations were based on previously published data<sup>20</sup>, which indicated an absolute difference in ejection fraction of 5% (with a pooled standard deviation of approximately 8%) between patients with an MoM hip prosthesis and those with a conventional hip prosthesis. Using an alpha value of 0.05 and a 2:1 sample size ratio between MoM and CoC prostheses, we needed 62 patients with an MoM implant and 31 with a CoC implant to have a power of 80% to detect a 5% difference (Cohen delta = 0.63, classified as a medium to large effect). The MoM group was divided according to blood metal levels using 7 ppb as a cutoff<sup>6</sup> (yielding approximately a 1:1:1 ratio overall).

### Statistical Analysis

The prespecified primary end points were ejection fraction (determined with CMR) and T2\* measurement. The secondary end points included left ventricular end systolic volume index, T1, and extracellular volume measurements with CMR; ejection fraction and left atrial area index and left ventricular long axis function (mean of the lateral and septal S' velocities measured with echocardiography); and blood biomarkers.

The 3 groups were assessed for matching using chi-square tests (or Fisher exact tests when the expected counts were <5) for binary variables and analysis of variance (ANOVA) for continuous variables. The distribution of the metal ions was not normal, and the group variances were highly heterogeneous; hence, nonparametric tests on medians were used to compare ion levels between groups. We used parametric and nonparametric test results to establish whether there was a significant difference in the mean and/or median effect on cardiac function between the groups. The distributions of the cardiac exposure variables in each group appeared normal for outcome variables of interest, with the variance usually

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constant (according to the Bartlett test); thus, ANOVA (F test) was conducted for each marker. As the sample sizes were relatively small, a formal test for normality was not conducted. However, sensitivity analyses with nonparametric tests (comparing the median across groups) were performed to relax the parametric assumption, and they always confirmed the conclusions obtained for the mean (results not shown). The presence of significant results at a 5% significance level was further assessed by dividing the threshold by the number of tests conducted (Bonferroni correction).

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Correlations between whole-blood metal ion levels and ejection fraction and T2\* values were calculated using the Pearson correlation coefficient (with metal ions log-transformed) and Spearman correlation coefficient (with metal ions on the original scale).

The minimum detectable effect size was calculated using Cohen's  $d^{28}$ .

All calculations were performed using Stata 14 (StataCorp).

## Results

### Patients

Ninety patients completed the study. Baseline patient characteristics are shown in Table I. Patients were matched for age and sex. Group A (CoC;  $n = 28$ ) had a mean age (and standard deviation [SD]) of  $65.3 \pm 8.8$  years, and 75% of the patients were female. Group B (MoM with a low metal-ion level;  $n = 33$ ) had a mean age of  $61.9 \pm 11.9$  years, and 64% were female. Group C (MoM with a high metal-ion level;  $n = 29$ ) had a mean age of  $67.6 \pm 10.8$  years, and 62% were female. The mean cobalt levels in groups A, B, and C were  $0.17 \pm 0.08$ ,  $2.47 \pm 1.81$ , and  $30.0 \pm 29.1$  ppb, respectively.

The overall mean time from implantation of the prosthesis to the date of the scan was slightly shorter in group A than in group B or C ( $84 \pm 33$  versus  $95 \pm 22$  and  $107 \pm 41$  months,  $F = 3.5$ ,  $p = 0.03$ ).

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Although the groups were not matched for body mass index (BMI), cardiovascular risk factors, or medications, there were no significant differences between them with regard to those factors.

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### CMR

The 3 groups did not differ significantly with regard to the mean left ventricular ejection fraction ( $70.1\% \pm 5.6\%$ ,  $69.5\% \pm 6.8\%$ , and  $70.6\% \pm 5.5\%$ ,  $p = 0.75$ ; Table II and Fig. 4) or the left ventricular end systolic volume index ( $p = 0.86$ ). Additionally, we found no differences in the left ventricular ejection fraction or end systolic volume index when we combined groups B and C and compared the combined group with group A (Table II).

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The mean cardiac T2\* values were normal in the 3 groups and did not differ significantly among them ( $p = 0.69$ ) or between the combined MoM group (B and C) and group A ( $p = 0.85$ ). All patients with high blood metal levels (Group C) had normal T2\* values ( $>20$  ms). The T1 values measured with both the MOLLI and the ShMOLLI techniques were normal in the 3 groups and did not differ significantly among the groups (MOLLI:  $p = 0.64$ ; ShMOLLI:  $p = 0.82$ ). The extracellular volume was normal in, and did not differ significantly among, the 3 groups ( $p = 0.28$ ), with no outliers.

The liver T2\* values were normal in all patients. No differences were identified among the 3 groups ( $p = 0.67$ ) or between the combined MoM group (B and C) and group A ( $p = 0.92$ ). The liver T1 value and extracellular volume also did not differ significantly between or among groups (Table II).

### Transthoracic Echocardiography

There was no significant difference in the left ventricular ejection fraction among groups A, B, and C ( $65\% \pm 6\%$ ,  $62\% \pm 7\%$ , and  $63\% \pm 6\%$ , respectively;  $p = 0.28$ ).

Similarly, the left atrial volume index was similar among the 3 groups ( $p = 0.19$ ) with no differences between the combined MoM group (B and C) and group A ( $p = 0.20$ ). There were also no between-group differences in the mean lateral and septal S' velocities ( $p = 0.46$ ).

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### **Blood Biomarkers**

There were no significant differences in the BNP level among groups A, B, and C (11.4, 10.8, and 24.9 pg/mL, respectively). Although the mean BNP level was highest in group C, the difference failed to reach significance ( $F = 1.17$ ,  $p = 0.32$ ). The troponin levels also did not differ significantly among the groups ( $p = 0.78$ ).

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### **Electrocardiography (EKG)**

All study participants had a resting 12-lead EKG. In 5 participants (4 in group B and 1 in group C), the EKG demonstrated left bundle branch block morphology (QRS duration of  $>120$  ms). There were no ST or T-wave abnormalities, or changes reflecting previous cardiac events such as myocardial infarction.

### **Metal Ions and Cardiac Function: Dose Response**

A dose-response linear correlation analysis was conducted using the Pearson coefficient to determine whether there was a relationship across groups between blood cobalt and chromium levels and left ventricular ejection fraction and T2\* values. There was no significant linear correlation between the blood cobalt level (log scale) and the left ventricular ejection fraction ( $R = 0.022$ , 95% confidence interval [CI] =  $-0.185$  to  $0.229$ ;  $p = 0.83$ ) or the T2\* value ( $R = 0.108$ , 95% CI =  $-0.105$  to  $0.312$ ;  $p = 0.32$ ). Similarly, no significant correlation was found between the blood chromium level and the left ventricular ejection fraction ( $R = 0.047$ , 95% CI =  $-0.162$  to  $0.251$ ;  $p = 0.66$ ) or the T2\* value ( $R = 0.065$ , 95% CI =  $-0.148$  to  $0.272$ ;  $p = 0.55$ ) (Fig. 5).

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Whole-blood metal levels were non-normally distributed, unlike the left ventricular ejection fraction and the T2\* value, and log-transforms attenuated without circumventing the lack of normality. Hence, to avoid this assumption, the Spearman correlation coefficient was also calculated, and it showed no significant correlation between the cobalt level and the left ventricular ejection fraction ( $\rho = 0.038$ , 95% CI =  $-0.170$  to  $0.243$ ;  $p = 0.72$ ), between the cobalt level and the T2\* value ( $\rho = 0.074$ , 95% CI =  $-0.139$  to  $0.280$ ;  $p = 0.50$ ), between the chromium level and the left ventricular ejection fraction ( $\rho = 0.058$ , 95% CI =  $-0.151$  to  $0.262$ ;  $p = 0.59$ ), or between the chromium level and the T2\* value ( $\rho = 0.012$ , 95% CI =  $-0.200$  to  $0.222$ ;  $p = 0.92$ ).

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### **Discussion**

There is increasing concern regarding the effects of systemic cobalt and chromium toxicity in patients with MoM hip implants. Although the evidence for toxicity is limited, patient and surgeon anxiety is fueled by increasing numbers of case reports of end-organ damage and mortality<sup>10</sup>. In this study, we investigated the effects of metal ions on cardiac function in 3 distinct groups of patients defined by the type of hip implant and the level of circulating blood cobalt. Using 2 independent cardiac imaging techniques (including the gold standard, CMR), we failed to find a significant change in cardiac function in patients exposed to elevated metal ion levels. There was no significant correlation between elevated blood metal ion levels and cardiac function; even at the extremes of the 95% CI around the correlation coefficients, the correlations were negligible to weak.

Our confidence in these results is based on the strengths of our study methodology, which included the use of both CMR and echocardiography as well as a meticulous trial design. We ensured a prespecified recruitment number, data acquisition in a dedicated cardiac imaging center separate from the recruiting center, and a cardiac care team

completely blinded to the study groups (during both acquisition and analysis) until the independent statistician returned the results.

Tissue-mapping techniques also failed to identify any demonstrable metal deposition from circulating cobalt ions in the cardiac or liver tissues. Although we recently demonstrated<sup>25</sup> that T2\* mapping can detect biopsy-proven CoCr, the lack of demonstrable tissue deposition in the present study may be due to differences in the physical chemistry of cobalt ions compared with that of cobalt and chromium in combination. However, the fact that iron can be detected in the heart and liver using T2\* mapping, and we previously detected CoCr (colocalized) in the liver<sup>25</sup>, indicates that the reason why the T2\* findings were negative in all cases in the present study was either that the test is too insensitive or no deposition was occurring. T2\* should be considered a candidate biomarker, which, if it had differed between the groups, would have indicated the biological process of deposition. The lack of functional consequences (demonstrated by complementary and independent imaging modalities [CMR and echocardiography] and 2 blood biomarkers) and the lack of positive T2\* findings, although not definitive, are reassuring.

A potential for cardiotoxicity from metal implants, as demonstrated by an inferior left ventricular ejection fraction, has been previously reported. However, these reports are limited to individual case reports and to studies with less rigorous methodology. For example, Prentice et al.<sup>20</sup> used echocardiography, which has a higher interobserver variability than CMR, and that may account for the difference between their and our findings. We conclude that individual case reports of heart failure (mainly in patients with extreme ion levels) are not likely to be “the tip of the iceberg” or to indicate an unrecognized epidemic of occult MoM-related community heart failure.

The pathophysiology behind a possible link between elevated blood cobalt levels and cardiac toxicity is unclear. Suggested theories include cobalt interference with cardiac myocyte oxygen uptake and transmembrane transport system disruption<sup>29</sup>. Histopathological findings in cobalt-related cardiac toxicity include myofibrillar hypertrophy, interstitial fibrosis, and muscle fiber degeneration<sup>30</sup>. However, calcified fibrils or other deposits within myofibrils are often absent, differentiating cobalt-induced cardiomyopathy from other etiologies<sup>13</sup>. It has been postulated that cobalt-related cardiac toxicity is due to additional predisposing factors, including poor nutrition and excessive alcohol intake as was seen in the Quebec beer-drinkers cobalt cardiomyopathy epidemic (in which foam stabilizer contained 10 times the usual quantity of cobalt)<sup>30</sup>. Recent case reports of cardiac toxicity in association with MoM hip implants mainly involved patients with extremely elevated blood cobalt levels (>100 ppb). Because of current national surveillance programs, such patients are now likely to have undergone revision surgery, with presumed reduction in their risk of systemic toxicity.

We believe that these findings offer reassurance to surgeons and will help them counsel the >1 million patients worldwide with MoM hip implants. Additional work is needed, particularly from large-volume linkage studies currently under way internationally.

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Fig. 1

Flow diagram of the study methods. ECHO = echocardiogram, LV = left ventricular, EF = ejection fraction, ESVi = end systolic volume index, ECV = extracellular volume, LAVi = left atrial volume index, BNP = B-type natriuretic peptide, and Trop I = troponin I.

Fig. 2

Comprehensive cardiac assessment included transthoracic echocardiography (**Fig. 2-A**), measurement of blood biomarker levels (BNP and troponin I) (**Fig. 2-B**), CMR (**Fig. 2-C**), and **scar** imaging (**Fig. 2-D**).

Fig. 3

Precontrast (**Fig. 3-A**) and postcontrast (**Fig. 3-B**) T1 mapping combined for extracellular volume mapping (**Fig. 3-C**).

Fig. 4

Scatterplot demonstrating the mean left ventricular ejection fraction (LVEF), and the SD, for the 3 groups.

Fig. 5

Correlation plots with line of best fit. Whole-blood cobalt and chromium levels are compared with the left ventricular ejection fraction (LVEF; top row) and T2\* values (bottom row).

Pearson correlation coefficients (R values) are provided for each.

**Commented [MD38]:** Please check. The text states "liver" and does not mention "scar". If "liver" is correct here, change "cardiac" to "cardiac and liver"?

TABLE I Patient Demographics by Study Group

Demographic	All Patients	Group A	Group B	Group C	P Value
Sample (no.)	90	28	33	29	
Age* (yr)	64.9 ± 10.5	65.3 ± 8.80	61.9 ± 11.9	67.6 ± 10.8	0.115
Sex (F/M) (no.)	60/30	21/7	21/12	18/11	0.525
Time since implantation* (yr)	8.76 ± 2.34	7.86 ± 2.62	9.03 ± 1.87	9.39 ± 2.52	0.041
BMI† (kg/m <sup>2</sup> )	28	28 (18-38)	28 (20-45)	27 (20-48)	0.929
Diabetes (no.; %)	3; 3	1; 4	1; 3	1; 3	0.982
Hypertension (no.; %)	32; 36	11; 39	12; 36	9; 31	0.715
Hypercholesterolemia (no.; %)	20; 22	9; 32	4; 12	7; 24	0.105
Medications (no.; %)					
β blocker	5; 8	2; 7	0	3; 10	0.181
ACE inhibitor‡	10; 11	5; 18	3; 9	2; 7	0.326
Thiazide diuretic	8; 9	2; 7	4; 12	2; 7	0.741
Calcium channel blocker	15; 17	7; 25	4; 12	4; 14	0.289
Aspirin	3; 3	1; 4	0	2; 7	0.317
Statin	20; 22	9; 32	4; 12	7; 24	0.121
Proton pump inhibitor	18; 20	8; 29	3; 9	7; 24	0.101
Thyroxine	8; 9	4; 14	4; 12	0	0.108
Hemoglobin level† (g/dL)	14	14.1	14.2	13.6	0.134
Cobalt level† (ppb)	10.9	0.17 (0.10-0.47)	2.47 (0.73-6.97)	30.01 (7.54-118)	<0.0001
Chromium level† (ppb)	7.7	0.74 (0.53-1.42)	2.84 (0.94-10.5)	19.6 (1.71-69.0)	<0.0001

\*The values are given as the mean and SD. †The values are given as the mean with or without the range in parentheses.  
 ‡ACE = angiotensin-converting-enzyme.

Commented [AB39]: 5/90 = 6%. Please check.

TABLE II CMR, Echocardiographic, and Blood Biomarker Results

Measurement†	Group A (N = 28)*	Group B (N = 33)*	Group C (N = 29)*	P Value	
				A vs. B vs. C	A vs. B and C
<b>CMR</b>					
Left ventricular ejection fraction (%)	70 ± 5	69 ± 7	71 ± 5	0.75	0.96
End systolic volume index (mL/m <sup>2</sup> )	21 ± 6	21 ± 8	20 ± 5	0.86	0.99
T1 MOLL: heart (NR = 949-1101 ms) (ms)	1,030 ± 42	1,014 ± 33	1,022 ± 37	0.64	0.15
T1 ShMOLL: heart (NR = 900-1,020 ms) (ms)	961 ± 31	957 ± 30	956 ± 44	0.82	0.54
T2*: heart (normal = >20 ms) (ms)	31 ± 5	31 ± 6	32 ± 6	0.69	0.85
Extracellular volume: heart	0.28 ± 0.03	0.27 ± 0.03	0.29 ± 0.04	0.28	0.62
T1 mapping: liver (ms)	616 ± 72	596 ± 78	584 ± 49	0.21	0.11
T2* mapping: liver (normal = >6.3 ms) (ms)	25 ± 4	26 ± 7	25 ± 5	0.67	0.92
Extracellular volume: liver	0.29 ± 0.04	0.30 ± 0.03	0.30 ± 0.04	0.62	0.36
<b>Echocardiography</b>					
Left ventricular end diastolic diameter (mm)	43 ± 9	45 ± 5	45 ± 6	0.26	0.10
Left ventricular end systolic diameter (mm)	28 ± 6	30 ± 4	30 ± 5	0.18	0.06
Left ventricular ejection fraction (%)	65 ± 6	62 ± 7	63 ± 6	0.28	0.12
Left ventricular lateral wall tissue Doppler imaging (m/s)	0.09 ± 0.02	0.08 ± 0.02	0.09 ± 0.02	0.24	0.16
Left ventricular septal wall tissue Doppler imaging (m/s)	0.08 ± 0.02	0.07 ± 0.02	0.08 ± 0.01	0.16	0.12
Left atrial volume index (mL/m <sup>2</sup> )	27 ± 6	28 ± 7	31 ± 8	0.19	0.20
<b>Blood biomarker sampling</b>					
BNP (normal = <47 pmol/L) (pmol/L)	11 ± 10	10 ± 8	25 ± 61	0.32	0.52
Troponin (NR = 0-14 ng/L) (ng/L)	7.38 ± 6.9	7.16 ± 5.0	8.62 ± 10.2	0.77	0.78

\*Data are presented as the mean and SD. †NR = normal range.

**Commented [MD40]:** Please add units (if any), here and for the liver ECV, and confirm that other units have been added correctly.

**Commented [AB41]:** Correct for TDI here and below?