1	Infectious causes of microcephaly: Epidemiology, Pathogenesis, Diagnosis, and					
2	Management					
3	Devakumar D*, Bamford A, Ferreira MU, Broad J, Rosch R, Groce N, Breuer J, Cardoso MA, Copp AJ,					
4	Alexandre P, Rodrigues LC, Abubakar I.					
5	*Corresponding author: Dr Delan Devakumar, UCL Institute for Global Health, 30 Guilford St,					
6	London. WC1N 1EH Tel: +44 (0)20 7905 2122 or +44 (0)7894 579082. <u>d.devakumar@ucl.ac.uk</u>					
7						
8	Citation: Devakumar D, Bamford A, Ferreira MU, et al. Infectious causes of microcephaly:					
9	epidemiology, pathogenesis, diagnosis, and management. Lancet Infect Dis 2017; published					
10	online Aug 22. http://dx.doi.org/10.1016/S1473-3099(17)30398-5					
11	http://thelancet.com/collections/infectious-diseases					
12						
13	Summary					
14	Microcephaly is an important sign of neurological malformation and predictor of future disability.					
15	The recent outbreak of Zika virus and congenital Zika infection has brought the world's attention					

_ _

16 to the links between infection and microcephaly. However, Zika virus is only one of the

17 infectious causes of microcephaly and, though the contexts in which they occur vary greatly, all

18 are of concern. In this review, we summarise important aspects of the major congenital infections

19 that can cause microcephaly, describing the epidemiology, transmission, clinical features,

20 pathogenesis and long-term consequences. We include the infections that cause substantial

21 impairment: cytomegalovirus, herpes simplex virus, rubella virus, Toxoplasma gondii, and Zika

virus. We highlight potential issues with the classification of microcephaly and show how some

23 infants affected by congenital infection may be missed or incorrectly diagnosed. While the

24 world's current focus on Zika virus is remarkable, preventing all infectious causes of

microcephaly and appropriately managing its consequences remain important global public health
 priorities.

27

28 Introduction

29 This review was conducted in response to the recent outbreak of microcephaly associated with congenital 30 Zika virus (ZIKV) infection, first identified in Latin America. Attention has been drawn to ZIKV because 31 the previously unknown congenital syndrome was discovered in a susceptible population and can 32 potentially affect a very large number of infants. ZIKV however is one of a group of infectious diseases 33 that can be transmitted to the fetus and cause microcephaly and the microcephaly is likely to represent the 34 tip of the iceberg in terms of developmental abnormalities, forming part of a syndrome for each infection. 35 Understanding the different causes of microcephaly will help clinicians, researchers, and hopefully policy 36 makers contextualize the recent outbreak and can be applied to help the individual child, as well as to 37 inform appropriate planning of services for the population. 38 In this article, we review the major congenital infections that can cause microcephaly, focusing 39 on those with the largest disease burden and the strongest evidence for causation: 40 cytomegalovirus (CMV), herpes simplex virus (HSV), rubella virus, Toxoplasma gondii (T. 41 gondii), and ZIKV. We describe the epidemiology, pathogenesis, transmission, clinical features 42 and long-term disability in childhood. Less frequent causes of primary microcephaly are not 43 considered here. While congenital syphilis has a high disease burden in regions affected by ZIKV 44 and varicella virus has recognised neurotropic properties, published reports describing primary 45 microcephaly in the context of these two infections are scarce.

46

47 Microcephaly

48 Microcephaly is a clinical diagnosis made at or after birth that describes a small head. It does not
49 necessarily mean abnormal brain development and some children with microcephaly are healthy.

50 An accepted definition of microcephaly is occipito-frontal head circumference (OFC)>2 standard 51 deviations below the median (<-2 z-scores), for gestational age and sex in an appropriate healthy 52 reference population, with severe microcephaly defined as <-3 z-scores.¹⁻³ The reference range 53 and population from which a definition is taken can vary, for example country-specific ranges, 54 the World Health Organization (WHO) child-growth ranges for term infants, the Fenton ranges 55 for preterm infants, and the InterGrowth reference ranges are all used.⁴ This limits the 56 comparability of microcephaly prevalence estimates and accounts for a proportion of the global 57 variability in rates. The WHO currently recommends the Intergrowth-21 criteria if the gestational age is known and the WHO Child Growth Standards if it is not.^{1,5,6} In addition, and probably 58 59 more importantly, inconsistency in frequency and rigour of measurement, and completeness of 60 reporting, makes population prevalences difficult to ascertain. A recent study across Europe 61 found a prevalence, excluding genetic conditions, of 1.53 (95% CI 1.16, 1.96) per 10,000 62 "births" (including live births, fetal deaths from 20 weeks gestation and termination of pregnancy 63 for fetal anomaly). Including genetic conditions would increase the prevalence to approximately 64 2.0 per 10,000. There was considerable variation in the estimates from 0.41/10,000 in Portugal to 65 4.25/10,000 in UK and in the definition of microcephaly (<-2 z-scores, <-3 z-scores and clinical 66 decisions), the reference ranges used and methods of ascertainment.⁷ Microcephaly can be divided into primary- i.e. develops before 32 weeks' gestation⁸ or birth⁹-67 68 and secondary- i.e. microcephaly that develops after this time. Primary microcephaly, the focus of 69 this review, is generally due to disturbed neurogenesis (mitosis or progenitor cell function) or death of neuronal progenitors.^{9,10} Secondary microcephaly normally relates to the postnatal 70

71 development and maturation of neurons (dendritic processes and synaptic connections).⁸ Infection

can lead to both primary or secondary microcephaly but this differentiation is often not possible if

the appropriate head circumference measurements are not taken. In high-income countries,

microcephaly is predominantly due to non-infectious causes,^{11,12} including genetic abnormalities,
nutritional deficiencies, hypothyroidism, brain injury, alcohol, drugs, and placental
insufficiency.^{8,9} A multi-centre retrospective analysis from Germany found that only 25 of 403
(6·2%) cases of microcephaly (for which a cause was identified) were due to maternal infections
in pregnancy.¹² The prevalence of in utero or perinatal infections in cases of severe microcephaly
is higher e.g. 46 of 284 (16%) cases in New York 2013-15.¹³

80 A reduction in head size may be associated with intrauterine growth restriction, for example with 81 CMV and ZIKV. This can be either symmetrically or asymmetrically reduced in size in relation 82 to the overall anthropometry of the infant. The gestation at which an insult occurs, will determine 83 when microcephaly is clinically detectable. An infection close to birth may result in brain damage 84 but with a normal head circumference. Recent data suggest that microcephaly may be poorly sensitive for screening infants severely affected by congenital ZIKV infection, particularly as this 85 consists of many other manifestations.¹⁴ Clinically apparent, microcephaly is one of many fetal 86 brain abnormalities that can lead to later neurodevelopmental sequelae.¹⁵ Although a clinical 87 88 diagnosis of microcephaly with normal brain imaging does not necessarily mean impaired brain growth, a reduction in skull volume is indicative of underlying cerebral cortical volume loss.^{8,16} A 89 90 study of Finnish infants admitted to a neonatal ward for example, showed that a one standard 91 deviation increase in head circumference at birth was associated with a 0.8-1.5 unit increase in cognitive scores at 56 months.¹⁷ 92

If we use microcephaly to screen for congenital diseases, and if OFC is normally distributed in the reference population, $2 \cdot 3\%$ of the population will be classified as having microcephaly, many of whom will be healthy. Conversely, many neonates with congenital infections that affect the brain will not have severe enough manifestations to cause microcephaly. To illustrate this, we simulate two scenarios (Figure 1). A definition of <-3 *z*-scores ($0 \cdot 1\%$ of the population)^{18,19} is

expected to be 99.9% specific, although much less sensitive (57%).⁴ An assessment of 1501 cases of suspected microcephaly reported to the Brazilian notification system, showed that 21.7% of notified cases classified as 'definite or probable' had a normal head circumference (>-2 z-scores) and this cut-off had a sensitivity of 83% and specificity of 98%.¹⁴ So although microcephaly is a good screening diagnosis for brain damage caused by a congenital infection, some cases will be normal and cases with neurological damage may not be identified. Further tools must therefore be employed.

105

106 Pathogenesis and embryology

107 The central nervous system becomes established from day 22 of embryonic development when the neural tube is formed.²⁰ The embryonic brain is initially composed entirely of proliferative 108 109 neuronal progenitors which reside within the ventricular zone (VZ) bordering the neural tube 110 lumen. However, with subsequent development, neurons begin to emerge and a new population 111 of deeper subventricular zone (SVZ) neural progenitors arises. Proliferation of the SVZ cells 112 contributes to further expansion of the brain's neuronal population. Proliferative VZ and SVZ 113 neuronal progenitors persist until around mid-gestation, providing a target for pathogens via the 114 cerebral blood supply which is now fully established. Experimental evidence shows that 115 regulation of neural progenitor numbers and sub-types is vital for controlling brain size and 116 morphology (Figure 2). For example, mouse studies reveal that abnormal expansion of the VZ or SVZ neural progenitors increases brain size and produces macrocephaly.^{21,22} In contrast, a 117 118 reduction in neural progenitor numbers due to cell death, cell cycle arrest or premature neuronal differentiation reduces brain size and produces microcephaly.²³⁻²⁶ 119

120

121 The pathogenesis of CMV, HSV, rubella and ZIKV is described below. Little information is

122	available on the mechanisms of congenital brain infection for T. gondii, where additional research
123	is needed. The congenital infections we describe in this paper (summarised in Table 1) are
124	responsible for a range of developmental brain defects, with the more severe cases occurring after
125	infection during the first trimester of gestation, when the neural progenitors are actively
126	multiplying and producing neurons (Figure 2). ²⁷⁻³⁶ Analysis of material from aborted human
127	fetuses confirms that brain cells are susceptible to viral infection (ZIKV, ³⁰ CMV, ³⁷ HSV ^{35,38} and
128	rubella ³⁹). Moreover, human cell culture systems demonstrate that neural progenitors are targeted
129	by these pathogens (ZIKV brain organoids ^{40,41} ; HSV iPS-derived cell culture ⁴² ; ZIKV
130	neurospheres ⁴³ ; CMV neural precursors ³⁷). This research suggests that perturbation of neural
131	progenitor populations may be the main cause of infection-related microcephaly.
132	
133	CMV
134	CMV is capable of altering progenitor and neuronal fates through the downregulation of
134 135	CMV is capable of altering progenitor and neuronal fates through the downregulation of multipotency markers including Sox2 and Nestin. ^{44,45} Neuronal differentiation has been found to
135	multipotency markers including Sox2 and Nestin.44,45 Neuronal differentiation has been found to
135 136	multipotency markers including Sox2 and Nestin. ^{44,45} Neuronal differentiation has been found to be inhibited or delayed by CMV ^{37,46,47} or to occur prematurely after infection. ⁴⁴ Recently, a new
135 136 137	multipotency markers including Sox2 and Nestin. ^{44,45} Neuronal differentiation has been found to be inhibited or delayed by $CMV^{37,46,47}$ or to occur prematurely after infection. ⁴⁴ Recently, a new molecular mechanism downstream of CMV brain infection has emerged: PPAR γ (peroxisome
135 136 137 138	multipotency markers including Sox2 and Nestin. ^{44,45} Neuronal differentiation has been found to be inhibited or delayed by $CMV^{37,46,47}$ or to occur prematurely after infection. ⁴⁴ Recently, a new molecular mechanism downstream of CMV brain infection has emerged: PPAR γ (peroxisome proliferator-activated receptor gamma) was found to increase following CMV infection of human
135 136 137 138 139	multipotency markers including Sox2 and Nestin. ^{44,45} Neuronal differentiation has been found to be inhibited or delayed by CMV ^{37,46,47} or to occur prematurely after infection. ⁴⁴ Recently, a new molecular mechanism downstream of CMV brain infection has emerged: PPARγ (peroxisome proliferator-activated receptor gamma) was found to increase following CMV infection of human neural stem cells and human fetal brain sections. Activation of PPARγ function alone was
 135 136 137 138 139 140 	multipotency markers including Sox2 and Nestin. ^{44,45} Neuronal differentiation has been found to be inhibited or delayed by $CMV^{37,46,47}$ or to occur prematurely after infection. ⁴⁴ Recently, a new molecular mechanism downstream of CMV brain infection has emerged: PPAR γ (peroxisome proliferator-activated receptor gamma) was found to increase following CMV infection of human neural stem cells and human fetal brain sections. Activation of PPAR γ function alone was sufficient to impair neuronal differentiation, and a PPAR γ inhibitor restored normal
 135 136 137 138 139 140 141 	multipotency markers including Sox2 and Nestin. ^{44,45} Neuronal differentiation has been found to be inhibited or delayed by $CMV^{37,46,47}$ or to occur prematurely after infection. ⁴⁴ Recently, a new molecular mechanism downstream of CMV brain infection has emerged: PPAR γ (peroxisome proliferator-activated receptor gamma) was found to increase following CMV infection of human neural stem cells and human fetal brain sections. Activation of PPAR γ function alone was sufficient to impair neuronal differentiation, and a PPAR γ inhibitor restored normal differentiation. Moreover, nuclear PPAR γ was detected in the brains of congenitally infected
 135 136 137 138 139 140 141 142 	multipotency markers including Sox2 and Nestin. ^{44,45} Neuronal differentiation has been found to be inhibited or delayed by $CMV^{37,46,47}$ or to occur prematurely after infection. ⁴⁴ Recently, a new molecular mechanism downstream of CMV brain infection has emerged: PPAR γ (peroxisome proliferator-activated receptor gamma) was found to increase following CMV infection of human neural stem cells and human fetal brain sections. Activation of PPAR γ function alone was sufficient to impair neuronal differentiation, and a PPAR γ inhibitor restored normal differentiation. Moreover, nuclear PPAR γ was detected in the brains of congenitally infected

to microcephaly is unclear. Infected organs including the brain, show tissue necrosis postmortem.³⁶ In vitro and animal studies suggest that HSV may initially induce an immune response
that stimulates neural stem cell proliferation. Subsequently, brain infiltration by CD8(+) T cells
limits proliferation through the stimulation of interferon-gamma.⁴⁹ Operation of a similar
immune-mediated mechanism following congenital brain infection leading to microcephaly is
possible but remains to be verified.

152

153 Rubella

The mechanisms by which rubella virus induces microcephaly remain largely unknown. Most human embryonic tissues can be infected by rubella virus^{39,50} and brain vessels have been found to degenerate following rubella infection.⁵⁰⁻⁵² This suggests that a neurodegenerative mechanism could be a potential underlying cause of rubella-induced microcephaly in humans. There is also indirect evidence that rubella infection can slow the rate of cell division although this has not been confirmed in human neural cells in culture.²⁸

160

161 ZIKV

162 Congenital viral infections and brain development studies have benefited from recent advances in 163 the growth of human cells in 3-dimensional (3D) culture. Cell aggregates called 'neurospheres' 164 are formed initially but with continued culture they can form 'organoids' that mimic some 165 features of true organs. Studies employing these techniques have confirmed that neural 166 progenitors (ventricular zone (VZ) and later subventricular zone (SVZ)) are indeed the commonest brain cell type infected by ZIKV (brain organoids^{40,41} and neurospheres⁴³). The 167 168 exposure of these 3D cell cultures to the virus mimics many features of microcephaly in humans, 169 including a decrease in neuronal production, reduced VZ thickness and overall smaller

organoids.^{40,41,53} This reduction in growth seems to result from cell cycle arrest⁵⁴ and/or an 170 171 increase in cell death, ^{41,43,53,54} and could indeed explain the microcephaly phenotype observed in 172 ZIKV-infected human fetuses. Entry of ZIKV into neural progenitors has been suggested to occur 173 via viral receptor AXL, which mediates ZIKV and dengue virus entry into human skin cells,⁵⁵ and is strongly expressed in VZ and SVZ neural progenitors.^{56,57} However, a recent study using 174 175 human brain organoids revealed that genetic ablation of AXL alone is unable to prevent ZIKV infection, suggesting that other cell adhesion or entry factors may be involved.⁵⁸ Human fetal 176 177 organotypic slice culture studies reveal that phospho-TANK Binding Kinase 1 (pTKB1), 178 relocates from centrosomes to mitochondria following ZIKV infection producing mitotic defects 179 and supernumerary centrosomes that may exacerbate cell death.^{57,59} Injection of ZIKV into 180 pregnant macaques and mice, or into the mouse brain, has recently confirmed some of the observations from human in vitro culture systems.⁶⁰⁻⁶² Dang and colleagues also discovered that 181 182 ZIKV infection activates TLR3 (involved in activation of an immune response) and that TLR3 inhibition can attenuate apoptosis and decrease growth induced by viral infection.⁴⁰ The 183 184 comparison of transcriptome profiles derived from infected and non-infected animal tissue also highlights new potential molecular viral targets. These include the down-regulated expression of 185 186 an extensive list of genes (all centrosome-related) previously associated with autosomal recessive primary microcephaly (ASPM, CENPF, MCPH1, STIL, Cep135).^{60,61} This suggests that the 187 188 underlying pathogenesis of autosomal recessive and virally induced microcephalies may be 189 similar, although this is yet to be confirmed.

190

191 Infection in pregnancy: Epidemiology, transmission and clinical features

192 Table 1 describes the epidemiology and clinical features of the main infections associated with

193 microcephaly in pregnant women/adults.

CMV

196	Most women of childbearing age in low- and middle-income country (LMICs) have long-lasting
197	immunity from prior CMV infection but viral reactivation can take place in pregnancy or during
198	periods of immunosuppression. In Sub-Saharan Africa, Latin America and South Asia, over 90%
199	of the general population have IgG antibodies compared to a seroprevalence of 40-60% in high-
200	income countries. ⁶³⁻⁶⁸ Prevalence of congenital CMV infection in high-income countries is
201	estimated to be 0.7% of all live births, or 1-5% in low-income countries. ^{63,66,69}
202	The risk of <i>in utero</i> transmission varies according to whether maternal CMV infection is primary,
203	a reactivation of latent infection, or superinfection with another CMV strain. ⁶³ Primary infection
204	has the highest risk of in-utero transmission and fetal disease is more severe, particularly when it
205	occurs earlier in pregnancy. Overall, most cases of congenital CMV infection result from non-
206	primary maternal infection but most of these are asymptomatic. ⁷⁰ The risk of transmission
207	increases with advancing gestational age, with 35% of mothers who have a primary infection in
208	the first trimester giving birth to infected newborns, compared to 65% who are infected in the
209	third trimester. ⁷¹ Rates of transmission in non-primary infection are less certain but are generally
210	reported to be lower. ^{63,72,73} The risk of congenital CMV infection is increased by maternal HIV
211	infection both for HIV exposed uninfected and HIV infected infants. ⁶³ In-utero HIV transmission
212	has been shown to be particularly associated with congenital CMV. ⁷⁴
212	

HSV

Primary microcephaly is a relatively rare complication of perinatal HSV infection and is mainly
found in association with in-utero infection which accounts for only 5% of cases.⁷⁵ Global

average seroprevalence is 18% amongst women of childbearing age but this includes huge
variations: from 4.1% in Japan to 62% in East Asia and 70% in Sub-Saharan Africa.^{76,77} Neonatal
infection at the time of birth from mothers with symptomatic genital infection is more common
but does not cause primary microcephaly.

222

223 Rubella

224 Rubella is a vaccine-preventable disease and rates of infection are largely dependent on the 225 coverage of immunisation programmes. The global incidence of congenital rubella syndrome has 226 reduced from 0.1-0.2 per 1000 live births prior to vaccination to near elimination (<0.01 per 100,000 live births) in areas with comprehensive vaccination rates.⁷⁸ Vaccination programmes 227 were widespread in most countries in 2014 except in Sub-Saharan Africa and South Asia.⁷⁹ 228 229 Despite widespread vaccination programmes an estimated 9.4% of pregnant women remain 230 seronegative worldwide and therefore susceptible to infection.⁸⁰ In areas without vaccination 231 programmes, incidence of congenital rubella syndrome has been estimated at 19-283 per 100,000 live births in the African region and 18-309 per 100,000 live births in the South Asian region.⁷⁸ 232 233 234 T. gondii 235 Most women of childbearing age in Latin America (51-72%), Central Europe (58%), and West Africa (54-77%) have specific IgG antibodies to *T. gondii*.⁸¹ Conversely, relatively few women of 236 237 childbearing age are found to be seropositive in South-east Asia, China and Korea (4-39%), Scandinavia (11-28%), and the United States (15%).^{81,82} Mother-to-child transmission 238 239 predominantly occurs following primary infection during pregnancy. The global incidence of 240 congenital toxoplasmosis is estimated at 1.5 cases per 1000 live births (95% credible interval 1.4, 241 1.6), being highest in some Latin America countries (average, 3.4 per 1000) and lowest in parts

of Europe (0.5 per 1000).⁸³ The risk of congenital infection may rise due to reactivation of *T*. *gondii* from immunosuppression, for example resulting from HIV.⁸⁴

244

245 *ZIKV*

ZIKV has been described for over 60 years with intermittent case reports until outbreaks in Yap
Island (estimated 73% (95% CI 68, 77) of population infected) in 2007, French Polynesia (up to
66% IgG +ve) in 2013 and Brazil in 2015.^{55,85,86} It is currently documented in 84 countries or
subregional areas and 31 have reported microcephaly or neurological malformations associated
with ZIKV.⁸⁷ The prevalence of suspected disease in women of childbearing age is 5400 per
100,000 in Brazil.⁸⁸

252

253 Laboratory diagnosis during pregnancy

Any pregnant woman presenting with fever and/or rash should be evaluated for risk of infections that are potentially transmitted to the fetus. There are many infectious causes of rash, for example Parvovirus B19, measles and varicella zoster, and extensive guidelines exist describing when testing should be done.⁸⁹

258 During the acute infection, laboratory confirmation is usually only possible where clinical 259 symptoms are present in the mother. Primary CMV, HSV, T. gondii, and ZIKV infections may be asymptomatic and therefore the opportunity to confirm acute infection can be missed.⁹⁰ In acute 260 261 symptomatic maternal infection, ZIKV and rubella can be detected in serum, blood, oral fluid or 262 urine by PCR amplification of nucleic acid and may precede the development of IgM antibodies. 263 Acute primary maternal HSV can be diagnosed by viral PCR and the presence of IgM antibodies 264 but usually only when oral or genital lesions are present. Virus may be detected for up 10 weeks in serum following acute ZIKV²⁹ and two weeks following acute rubella⁹¹ while CMV and HSV 265

are frequently shed asymptomatically from mucosal epithelia.^{92,93} Detection of *T. gondii* IgA is the most sensitive indicator of congenital infection in the child.⁹⁴ CMV shedding in oral fluid may occur both during primary infection and asymptomatic reactivation and is therefore not always a reliable indicator of acute primary infection. Detection of virus needs to be interpreted in the light of serological results.⁶⁶

271

272 Serological testing in pregnant women

273 For rubella, T. gondii and CMV commercial enzyme immunoassays are routinely available for 274 antibody detection. For rubella and toxoplasmosis cases, serological diagnosis of acute primary 275 infection relies on detection of acute phase IgM antibodies, which for most do not reach adequate 276 levels until one week following acute infection. For all three, paired IgG samples taken at least 277 10-14 days apart should also be obtained to confirm seroconversion. A fourfold rise in antibody titre is indicative of recent although not necessarily primary infection.⁹⁴ T. gondii IgM antibodies 278 279 are commonly detected by ELISA in acute infections, but may persist for several months.⁹⁵ High levels of IgG antibodies may also be present during the acute phase of infection.⁹⁶ 280 In general, detection of CMV IgM, has been shown to lack sensitivity and specificity. especially 281 282 in conditions of immune dysfunction. A positive CMV IgM is associated with primary infection 283 in only 10% of cases. To distinguish between acute and chronic infections, ELISA-based IgG 284 avidity tests are widely recommended: low-avidity antibodies are typically found over the first 285 weeks, while high-avidity IgG predominates in the chronic phase.⁹³ 286 IgM and IgG antibody tests for dengue, West Nile fever and ZIKV members of the flaviviridae

287 family share considerable epitope cross-reactivity. Plaque reduction neutralising tests can be used

to differentiate closely related viruses but is too complex for routine diagnosis especially in non-

289 specialised laboratories.

290

291 In-utero testing

292 In the event that acute maternal infection with any of the above pathogens is considered to pose a 293 risk, infection of the infant can be investigated where possible by amplification of pathogen 294 nucleic acid from amniotic fluid. Amniotic fluid samples are typically obtained at 18 weeks gestation to determine fetal infection with T. gondii and guide therapy.⁹⁷ In CMV infection, there 295 296 is a 6-8 week window between maternal infection and being able to detect the virus in amniotic 297 fluid. PCR has high sensitivity when performed at the correct time (20-21 weeks gestation or 7 298 weeks after maternal infection) and, when combined with culture, nearly all congenital infection 299 can be diagnosed.⁶³ PCR testing of amniotic fluid for HSV is possible but does not seem to correlate with neonatal infection,⁹⁸ and has recently been shown to be possible with ZIKV.⁹⁹ 300 301

302 Screening

303 Childhood immunisation programmes against rubella are routinely available in 147 countries, with an estimated global coverage of 46%.¹⁰⁰ Pregnant women may be screened for rubella 304 305 antibody and those in whom levels are absent or low, offered post-partum vaccination. Recently, 306 high coverage levels of immunisation have led to some high-income countries to drop rubella 307 screening. Prenatal screening for toxoplasmosis is routinely undertaken in Austria, France and Slovenia and neonatal screening in Denmark, Ireland and some parts of USA and Brazil.⁹⁵ In 308 309 France, women who develop high titres of IgM antibodies or evidence of seroconversion 310 indicative of primary infection during pregnancy are offered fetal screening and treatment with 311 spiramycin or pyrimethamine-sulphonamide. Severe sequelae from congenital toxoplasmosis are 312 now rarely seen after the current approach of systematic prenatal screening and treatment was implemented in France.¹⁰¹ Screening for CMV in some high-income countries and for ZIKV in 313

314 countries with high levels of transmission are being considered however, it is not currently315 recommended for either.

316

317 Clinical presentation at birth

Whilst infection can cause both primary and secondary microcephaly, the main focus of this review is the effect of congenital infections on neurogenesis or antenatal neural progenitor death usually associated with primary microcephaly, i.e. it is present at birth. Evaluation of an infant with suspected microcephaly at birth seeks to meet three goals: (1) confirm the diagnosis of microcephaly; (2) identify the cause and attempt syndromic diagnosis; and (3) aid prognosis and guide initial treatment where appropriate.

324 Measurement of the OFC (illustrated in Figure 3) is used as a screening examination to identify 325 neonates who are likely to have an underlying neurological condition. A full antenatal history 326 with particular focus on acquired environmental insults, maternal serology, antenatal fetal growth 327 measurements where available, and family history of microcephaly and neurological conditions 328 are essential to identify possible causes. Systemic examination, including ophthalmology and 329 audiology, will help in identifying associated abnormalities in other organ systems. Cranial 330 ultrasound (crUS) can be used to screen for anatomical abnormalities where available. Features 331 suggestive of congenital infections in an infant with microcephaly are: maternal history of 332 infection during pregnancy, indicative antenatal maternal serology results, clinical features in the 333 neonate (Panel) and calcifications on brain imaging. In patients with these features, a more 334 specific work-up is required, including serological testing to confirm specific infectious causes, other laboratory investigations, and further neuroimaging. 335

336

337 Identifying Specific Infectious Causes

Common clinical features for each infection are shown in the appendix and a summary of
diagnosis and treatment recommendations are given in Table 2. Much of this information is based
on historical cohorts and case studies and is therefore potentially prone to bias. Individual patients
may present with different signs and symptoms.

342

343 CMV

344 Most neonates with congenital CMV are asymptomatic but common manifestations in

345 symptomatic neonates include intra-uterine growth restriction, sensorineural hearing loss,

346 petechiae, and jaundice. Neurological sequelae are observed in 60-90% of those with clinical

347 symptoms at birth,¹⁰² although biases from early cohort studies may overestimate this.^{72,103,104} Up

to 15% of infants that are asymptomatic at birth may go on to develop symptoms later in

349 childhood.^{63,102,104}

350

351 *HSV*

352 Infection can occur *in utero*, intrapartum or postnatally. *In utero* infection is classically associated

353 with a triad of cutaneous, ophthalmological and neurological abnormalities (including

354 microcephaly). Vesicular lesions may or may not be present and often develop late. Perinatal

355 infection presents in 3 main ways: disseminated disease affecting multiple sites, CNS disease or

356 skin/eyes/mouth limited infection.^{75,105}

357

358 Rubella

Fetal abnormalities resulting from *in utero* transmission of rubella range in severity according to the gestation at the time of infection. Infection in the first trimester is associated with more severe abnormalities.¹⁰⁶ Symptomatic infection in the infant is referred to as congenital rubella syndrome (CRS) with a classic triad of cataracts, sensorineural deafness and cardiac defect (e.g. patent
 ductus arteriosus, ventricular septal defect).¹⁰⁷ Estimates of the frequency of microcephaly in
 CRS vary but have been reported to be as high as a third of cases overall.¹⁰⁸

365

366 T. gondii

Approximately 24% of live born infants infected with *T. gondii* are symptomatic at birth.¹⁰⁹ The 367 368 classic signs originally described by Sabin (chorioretinitis, microcephaly or hydrocephalus, and 369 widespread intracranial calcifications) are relatively infrequent, but highly suggestive of the 370 diagnosis of congenital toxoplasmosis. More severe manifestations occur in infections earlier in 371 gestation. Intracranial lesions, for example, are seen in up to 40% of congenital infections acquired before 5 weeks of pregnancy but in less than 10% of those acquired beyond 20 weeks.¹⁰⁹ 372 12.5% of liveborn infants with congenital toxoplasmosis develop severe neurological sequelae;¹¹⁰ 373 374 5% of them have microcephaly.¹¹¹

375

376 *ZIKV*

ZIKV infection in pregnancy and its possible link to a range of birth defects is currently the 377 378 subject of intense investigation worldwide. There is now convincing evidence that ZIKV 379 infection in pregnancy, especially in the first trimester, is associated with an increased risk of 380 microcephaly. The risk of microcephaly associated with ZIKV infection was estimated as 95 per 381 10,000 infected women in first trimester in French Polynesia.¹¹² Congenital disease has 382 predominantly been seen in Brazil, but this may spread across Latin America. Brasil et al 383 followed up ZIKV affected pregnant women in Rio de Janeiro and found that 3.4% had 384 microcephaly and 42% had abnormal clinical or radiological findings in the first month, mostly affecting the central nervous system.¹¹³ The spectrum of congenital Zika syndrome (CZS) 385

386 includes birth defects associated with microcephaly (including hearing loss and ophthalmological 387 defects) and how they are related to timing of infection, and presence or absence of maternal 388 symptoms, as well as the influence of other arboviral infections has yet to be fully delineated. 389 Currently, most of what is known about CZS comes from neonates with microcephaly which 390 appears to include syndrome-specific features such as partially collapsed skull, reduced cortical thickness and extensive subcortical calcifications.^{114,115} However, the full spectrum will only 391 392 become clear when cohorts of infected mothers provide in-depth report of the clinical 393 presentation. Brain damage or ocular lesions can be found in the absence of microcephaly and 394 other features, as described in the appendix, are common.^{14,116,117} It is yet to be shown whether 395 there is ongoing viral replication at the time of birth in congenital ZIKV infection. This will have 396 important implications for the potential efficacy of postnatally administered antiviral therapy. 397

398 Markers for Prognosis

399 Microcephaly outcomes are varied, and more accurate prognosis at the time of diagnosis is one of 400 the main aims of clinical evaluation of the neonate. Neuroimaging- for example crUS, CT and 401 MRI- has proven a useful predictive investigation and is warranted in neonates with severe 402 microcephaly, or where a congenital infection is suspected.¹¹⁸ Whilst different modalities have 403 specific advantages (e.g. good visibility of calcifications on CT, bedside availability of crUS), 404 many structural abnormalities caused by disruptions in development can be identified across 405 modalities. Typical abnormalities include intracranial calcifications, white matter abnormalities 406 (e.g. periventricular leukomalacia, delayed myelination; best seen on MRI), gyration defects (e.g. 407 polymicrogyria), and schizencephaly.^{12,119}

408

409 Long-term consequences and resulting disability in childhood

410 Long-term follow-up is recommended for infants with microcephaly and even for those with 411 congenital infections and who are apparently unaffected at birth in order to manage evolving 412 conditions and identify new manifestations early. In many cases, the cause of microcephaly will 413 not be known and generic plans can be adopted to manage impairments and limit disability. The 414 timing of the interventions vary in importance. Hearing screening for example needs to be done 415 early to enable interventions to optimise language development. Other interventions, including psychosocial support and counselling,¹²⁰ are required throughout childhood and into adult life. 416 417 Where microcephaly occurs, recommendations are for early intervention to address associated impairments, calling upon a host of specialist medical and educational services.¹²¹ Currently, 418 419 regular follow-up is recommended for ZIKV associated microcephaly for anthropometry, 420 developmental and neurological assessments and also hearing and ophthalmological assessment 421 where required over the first two years, with less frequent follow-up recommended for infants 422 without microcephaly.¹ Evidence-based therapy strategies exist that may improve cognitive outcomes and reduce disability.¹²² Interventions in early life, for example child stimulation,¹²³ 423 424 maximise the plasticity of the developing brain, attenuating the consequences of the damage to 425 the nervous system. But such recommendations ignore the realities of life in many countries. 426 Issues begin at birth, for example, in some societies, children with visible disabilities are allowed 427 to die in the neonatal period, either through active infanticide or through withdrawal of basic care such as feeding.¹²⁴ 428

The provision of both recommended early diagnosis, and appropriate interventions may be severely limited in countries with only a handful of specialists, where early intervention programmes or inclusive education efforts remain rare and are largely urban-based. Based on a small but growing literature on disability, individuals with more severe disabilities are far less likely to receive appropriate medical care, attend school or are included in the social, economic or religious life of their communities.¹²⁵ This is compounded in poorer households, which often
choose to invest limited resources on non-disabled children, whom they feel will be able to
contribute to the household in future.¹²⁶ Furthermore, recommendations for individuals with
microcephaly tend to concentrate on early childhood, but anticipate little about their management,
support or advocacy needs beyond pre-school years. While attitudes vary from society to society,
overwhelmingly, persons with disabilities face increased risk of stigma, social isolation, abuse
and poverty across their lifespan.¹²⁷

Infections that cause microcephaly are likely to disproportionately affect LMICs and within these countries, the poorest populations, who are more likely to live in crowded areas with inadequate housing and limited water and sanitation systems. Compounding this, poorer women are less likely to have access to family planning services, to access prenatal screening or, if needed, safe termination of pregnancy.¹²⁸

Although social protection schemes are beginning in many middle-income countries,¹²⁹ the cost of raising and supporting a disabled child continues to be borne almost entirely by the immediate family. Households with disabled members are on average, poorer because of increased costs and because family members, in particular women, must take time away from income generating activities to provide care.¹³⁰

451 Improved services and support for affected individuals and families across the lifespan must be

452 anticipated, and health professionals must work with civil society organisations, including

disabled peoples organisations, to ensure these children and their families receive the medical,

454 educational and social service support they need and are entitled to.¹³¹

455

456 Conclusions

457 We have summarised the epidemiology, clinical presentation and the current understanding of the

458 pathogenesis of the major congenital infections associated with microcephaly. Potential 459 inconsistencies in the criteria used to diagnose microcephaly have been highlighted as well as 460 limitations of current diagnostic methods in confirming an infectious aetiology. If microcephaly, 461 as strictly defined by head circumference, is used as a screening tool, it will miss affected infants 462 who do not manifest with a small head size. Many infants are likely to have subtle deficits 463 needing more sensitive neurological and developmental assessments. Microcephaly is also 464 usually only one manifestation in a syndrome and other assessments are required to identify the 465 full spectrum of illness.

466 ZIKV has brought the attention of the world to the problem of microcephaly. The extent of the 467 disease burden resulting from ZIKV is still being clarified but it appears to be significant and is 468 one of a number of infections associated with primary microcephaly. On an individual level, 469 accurate diagnosis and thorough investigation of confirmed microcephaly can guide management 470 and determine prognosis. On a population level, assessing the overall burden and contribution of 471 different causes will guide future interventions and strategies for prevention. Preventing infection, 472 where possible, must be prioritized. It is not within the scope of this review to fully consider 473 health protection and primary prevention but key interventions exist, for example population 474 immunisation for rubella and vector control for ZIKV. Pharmacological interventions are able to 475 treat some congenital infections and therefore limit further damage to the child's developing 476 nervous system (CMV, HSV, T.gondii). Antiviral treatments are not currently available to treat 477 ZIKV in pregnancy but vaccines are in development. Interventions to attenuate the effects of the 478 developmental delay, and at the community or population level to minimize impairment caused 479 by infections that can lead to microcephaly are all needed. More clinical and health system 480 research is crucial to provide scalable interventions to reduce the disability associated with 481 microcephaly. Many interventions designed to optimise neurodevelopmental outcomes in

482	infected infants will be common to all congenital infections and enhanced services for ZIKV				
483	follow up could potentially also be used in the future for the benefit of children severely affected				
484	by other congenital infections.				
485	Regions in which the infective burden is highest are also likely to be those where diagnosis and				
486	management is most lacking, creating a disproportionately large burden on populations that can				
487	least afford it. Inequities in access to optimised multidisciplinary care for children with				
488	neurodisability in areas with the highest rates of microcephaly means that the prognosis for those				
489	infants is often very poor. ZIKV has highlighted the issue of microcephaly and provides a timely				
490	opportunity to refocus on the whole group of infectious diseases which can affect the developing				
491	fetal brain and to redouble our efforts for prevention and treatment in the short- and long-term for				
492	those living with the frequently devastating consequences of such congenital infections.				
493					
494					
495	Panel: Clinical features suggestive of congenital infection in a neonate with				
496	microcephaly. ^{132,133}				
497	• Severe microcephaly (<-3 <i>z</i> -scores)				
498	• intrauterine growth restriction				
499	hydrops fetalis				
500	• seizures				
501	• cataract and other visual abnormalities				
502	hearing loss				
503	congenital heart disease				
504	hepatosplenomegaly				
505	• jaundice				

506 • characteristic rashes

Organism	Description	Transmission	Risk period for	Maternal signs and	Prevention of infection and
			transmission of infection	symptoms	treatment of pregnant
			to the fetus		women
CMV	Betaherpes DNA	Infectious body fluid (saliva,	Risk increases with	Mostly asymptomatic or	Treatment with antiviral
	virus	urine, breast milk, genital	gestational age, but earlier	flu-like symptoms	medication not currently
	Herpesviridae	secretions, blood transfusion).	infection is associated with		recommended outside of
	family		more severe congenital		trial settings.
			infection.		Candidate vaccines are in
					development. Hyperimmune
					globulin has been trialled but
					has not as yet been shown to
					reduce transmission of CMV
					to the fetus. ¹³⁴
HSV 1 & 2	Alphaherpes	Orogenital, genital-genital,	Risk is associated	Primary genital herpes	Risk of perinatal
	DNA viruses.	transmitted during delivery	predominantly with	infection may be	transmission can be reduced
	Herpesviridae	through infected maternal	primary infection late in	asymptomatic but is often	by Caesarean section in the
	family	genital tract.	pregnancy.	associated with	context of active genital
		Virus shed with or without		mucocutaneous lesions. ⁷⁵	lesions and suppressive

Table 1. Summary of the microbiological, virological and maternal clinical features

		lesions present.		Latent infection is	aciclovir/valaciclovir given
				subsequently established in	to the mother prior to
				dorsal root ganglia.	delivery. ¹³⁵
				Mucocutaneous lesions can	
				then recur following viral	
				reactivation. ¹⁰⁵	
Rubella virus	Single stranded	Respiratory transmission.	Risk of congenital disease	Typically a mild self-	Immunisation of girls to
	RNA virus	Humans are the only known	in first 20 weeks; infection	limiting illness	prevent individual cases and
	Togaviridae	host of rubella. ¹³⁶	beyond this not associated	Maculopapular rash,	the whole population to
	family		with congenital defects.	lymphadenopathy, malaise,	interrupt transmission.
			Primary maternal infection	arthralgia, fever. ¹³⁶	Supportive management of
			is associated with up to a		the infection.
			50% risk of fetal		
			infection. ¹³⁷		
T. gondii	Obligate	Humans acquire infection	Risk of congenital	Most acute <i>T. gondii</i>	Treatment with spiramycin
	intracellular	mainly by consuming raw or	transmission increases with	infections in	and/or pyrimethamine-
	protozoan	undercooked meat containing	gestational age at maternal	immunocompetent	sulphonamide of maternal
	parasite	cysts, by ingesting water or	infection: 15% (95% CI	pregnant women are	toxoplasmosis diagnosed
		raw vegetables contaminated	13–17) at 13 weeks and	subclinical; fever,	through prenatal screening is
		with oocysts, or by	71% (95% CI 66-76) at 36	lymphadenopathy and a	believed to reduce the risk of

	tachyzoites during an acute infection. Tachyzoites invade host cells, multiply and		common clinical signs and symptoms. ¹³⁸	and neurologic sequelae in the fetus in settings where
	host cells, multiply and		symptoms. ¹³⁸	the fetus in settings where
				low-virulence type II strains
	disseminate, infecting			predominate, but this
	multiple sites in the body			protective effect remains to
	including brain, eye, heart,			be confirmed against more
	skeletal muscle and			virulent atypical
	placenta. ¹³⁸			recombinant strains. ¹⁰¹
	Toxoplasmosis reactivation in			
	immunocompromised women			
	may rarely be a cause. Blood			
	transfusion or organ			
	transplantation.			
	The only definitive hosts are			
	members of the cat family. ¹³⁸			
ngle stranded	The main vector is the <i>Aedes</i>	Risk of transmission	Infection is often	Primary prevention of
NA arbovirus	mosquito. A. aegypti is	appears to be throughout	asymptomatic in adults	mosquito bites. No treatment
aviridae	considered of greatest global	pregnancy	with symptoms reported in	as yet.
	importance. ¹³⁹		approximately 20% of	
N	A arbovirus	skeletal muscle and placenta. ¹³⁸ Toxoplasmosis reactivation in immunocompromised women may rarely be a cause. Blood transfusion or organ transplantation. The only definitive hosts are members of the cat family. ¹³⁸ gle stranded The main vector is the <i>Aedess</i> mosquito. <i>A. aegypti</i> is considered of greatest global	skeletal muscle and placenta.138Toxoplasmosis reactivation in immunocompromised women may rarely be a cause. Blood transfusion or organ transplantation.The only definitive hosts are members of the cat family.138gle strandedThe main vector is the Aedes mosquito. A. aegypti is considered of greatest globalpregnancy	skeletal muscle and placenta.138skeletal muscle and placenta.138Toxoplasmosis reactivation in immunocompromised women may rarely be a cause. Blood transfusion or organ transplantation

provisional)	family	Also sexual transmission and	cases. ¹⁴¹	
		potentially through infected	Common features include	
	There are	blood transfusion.140	pruritic, maculopapular	
	currently two	Non-human hosts are thought	rash, low grade fever,	
	major lineages:	mainly to include non-human	arthritis/arthralgia,	
	one African	primates, although evidence	conjunctivitis, myalgia and	
	(with two	of infection has also been	headache.	
	groups, the	demonstrated in other	Has been associated with	
	Uganda cluster	mammals. ¹³⁹	Guillain-Barré syndrome,	
	and the Nigeria		myelitis and	
	cluster) and one		meningoencephalitis	
	Asian/American.		reported in adults. ¹⁴²	

Table 2: Diagnosis and treatment of infants

Organism	Diagnosis	Treatment	
CMV	CMV viral culture or PCR testing for DNA from urine or saliva	Early treatment of confirmed symptomatic congenital CMV with IV	
	obtained within the first three weeks of life. Retrospective diagnosis	ganciclovir or oral valganciclovir is currently recommended based	
	can be achieved from dried blood samples taken for newborn	on the results of 2 main trials. ¹⁴³⁻¹⁴⁵ Results from randomised trials	
	screening. Testing should be done as early as possible.	and observational studies have demonstrated improved hearing and	

		neurodevelopmental outcomes although questions remain regarding
		optimal treatment strategies, including when to treat and duration of
		therapy.
HSV 1 & 2	PCR testing for viral DNA on surface swabs (conjunctivae, mucosal	Intravenous high-dose acyclovir is indicated for confirmed neonatal
	surfaces), cutaneous lesions, CSF, whole blood, and tracheal	HSV infections. ¹⁴⁶
	secretions where available.	
Rubella	Isolation of rubella virus (or viral RNA) from the neonate, isolation	Management is mainly supportive and involves monitoring for late
	of rubella IgM or persistent rubella IgG. The virus is most	emerging symptoms (e.g. hearing loss, endocrine problems).
	commonly isolated from nasopharyngeal samples, but can be from	
	blood, urine, and CSF cultures. ¹⁴⁷	
T. gondii	Diagnosis is difficult if specific IgM and IgA antibodies are not	Congenital toxoplasmosis is treated with pyrimethamine and
	detected in the serum or plasma at birth. CSF samples to detect IgM	sulphonamide (sulphadiazine or sulphadoxine) with folinic acid to
	and IgA antibodies may confirm the diagnosis in infants with	minimize pyrimethamine-associated hematologic toxicity. Up to
	intracerebral lesions.	85% of children with subclinical congenital infection, if left
	Maternal IgG is detectable in the fetus for several months, generally	untreated, will later develop signs and symptoms of disease, such as
	disappearing completely within one year. Because specific IgG	chorioretinitis or developmental delays. ¹⁴⁸
	produced by congenitally infected newborns may be detected about	
	12 weeks after birth, the presence of high IgG titres beyond this time	
	is suggestive of congenital infection. ⁹⁵	

ZIKV	Testing for IgM antibody with capture ELISA using recombinant	Clinical management of complications, including dysphagia,
	antigens in CSF or blood at birth indicate congenital infection;	irritability and epilepsy. Supportive care for neurocognitive delays,
	plaque reduction neutralisation tests are required to confirm	hearing and visual loss, and appropriate developmental follow-up.
	monotypic antibody responses, although dengue and yellow fever do	
	not cause congenital infections. Reverse-transcriptase PCR detects	
	acute infections, positivity rare in neonates. Tests can be done on	
	serum/plasma from cord blood or infant's peripheral blood. IgM	
	detection in CSF of neonate is highly specific.	

CNS = central nervous system, CSF = cerebrospinal fluid, PCR = polymerase chain reaction

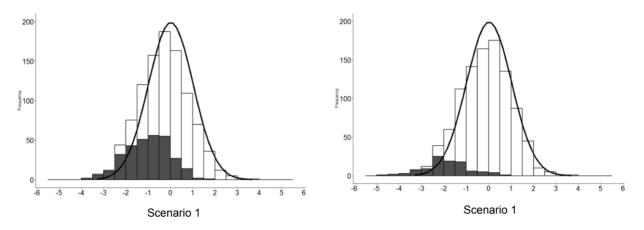


Figure 1: Modelled population effects of insults to brain developmental

* Bars represent the OFC distribution at birth (*z*-scores on *x* axis). The dark bar segments indicate the head circumference measurements of these "affected" children. The continuous line represents the reference population (no children "affected"), with a mean of 0 and SD of 1. Simulated populations were created using R language and environment for statistical computing.¹⁴⁹

If an otherwise normal population of 1000 infants (in which children's occipito-frontal circumferences (OFCs) are close to their expected values) is exposed to a congenital infection that limits brain development, a proportion of children are labelled as "normal" by being above the -2 *z*-scores (standard deviations (SD)) or -3 *z*-scores cut-offs but they may deviate from their "expected" OFC.

Scenario 1 describes a congenital infection with a uniform shift to the left by 1 SD relative to the reference curve in 30% (randomly chosen) of affected children, roughly as expected in congenital toxoplasmosis acquired late in pregnancy. The remaining 70% children were not affected at all. In this hypothetical scenario, 6.5% of children fall below -2 *z*-score cut-off value for microcephaly; 0.9% below the -3 *z*-scores. Only 65 of the 300 (21.7%) of the "affected" children will actually be classified as "microcephalic" by using the -2 *z*-score cut-off. Only children whose "ideal" OFC would be less than -1 SD of the expected mean (expected proportion of 15.9% in a

healthy population), would actually be diagnosed as "microcephalic" after the 1 SD shift caused by the infection.

Scenario 2 is a congenital infection where OFC is shifted to the left by 2 SD in 10% of children, roughly as expected for congenital toxoplasmosis acquired early in pregnancy. In this hypothetical scenario, 6.5% of children fall below -2 *z*-score cut-off value for microcephaly; 1.4% below the -3 *z*-scores. Here, 65% of "affected" children will fall below the cut-off value of -2 SD.

By counting the number of children below a given cut-off value we are severely underestimating not only the proportion of children who have been relatively mildly affected (Scenario 1) but also that of children who have been more severely affected (Scenario 2). Moreover, although Scenarios 1 and 2 represent conditions varying in severity which affect different proportions of children in the population, in both examples the prevalence of microcephaly, estimated by applying the -2 *z*-score cut-off, would be quite similar (around 6.5%).

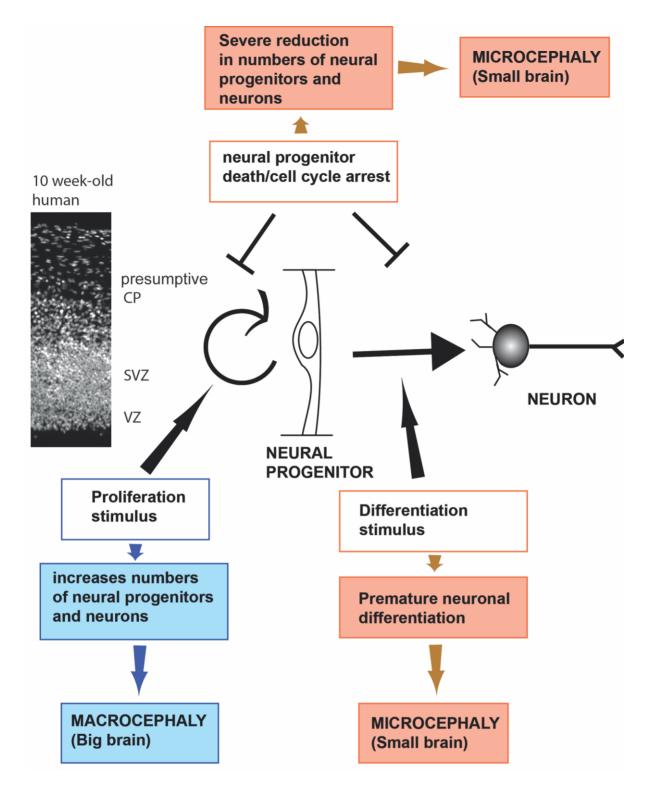


Figure 2. Factors that may influence neuronal and neural progenitor populations and lead

to microcephaly or macrocephaly

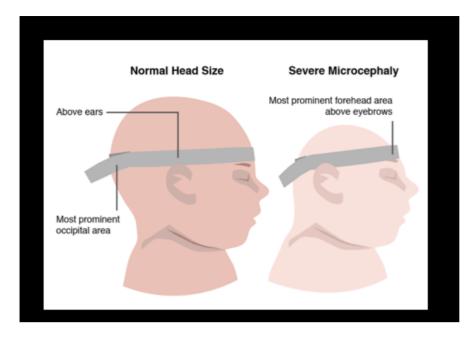


Figure 3: Measurement of occipito-frontal circumference (OFC) in the neonate

Measurements of OFC can be variable and user dependent. To aid robust measurements, a nonstretchable tape should be used and placed above the ears, covering the broadest part of the forehead, and the most prominent area of the occiput as shown. The largest measured circumference of three repeat measures should be used and recorded to the nearest millimeter. In neonates, the most robust measurements are achieved at >24h of age when post-partum skull modelling has subsided.

Search strategy

Pubmed, Embase and Google Scholar were searched, with no language or date restrictions (search end date: 31 March 2017), for the following terms and reference lists were searched for additional citations:

• (Cytomegalovirus or CMV or Rubella or MMR or Zika or HSV or herpes or toxoplasma or toxoplasmosis) and (seroprevalence or prevalence or incidence)

- (congenital pathogenic infections or pathogenic infections) and (brain disorders or microcephaly)
- (pathogens or virus or pathogen or viral) and microcephaly
- (Cytomegalovirus or CMV or Rubella or MMR or Zika or HSV or herpes or toxoplasma or toxoplasmosis) and (brain or brain disorder or brain development or brain defects)
- (cytomegalovirus or rubella or toxoplasmosis or herpes simplex or varicella or chikungunya or West Nile virus or HIV or syphilis) AND microcephaly NOT Zika

Conflicts of interest

We declare that we have no conflicts of interest.

Acknowledgements

Fabien Lafaille who advised on HSV and brain development. Joint MRC/Wellcome Trust (099175/Z/12/Z) Human Developmental Biology Resource (<u>www.hdbr.org</u>) for providing a human brain section shown in Figure 2.

Author contributions

DD and IA conceived the work. AB, DD, JoB, MUF, PA and RR undertook the literature reviews. AB, AJC, DD, JoB, JuB, LCR, MAC, MUF, NEG, PA and RR wrote the sections of the draft. All authors interpreted and critically revised the draft.

Funding

There was no specific funding for this work. DD receive salary support from NIHR, and PA from Royal Society, Dorothy Hodgkin Fellowship. IA is supported by NIHR (SRF-2011-04-001; NF-SI-0616-10037). JuB receives funding from the NIHR UCL/UCLH Biomedical Research Centre.

LCR is partially funded by the European Union's Horizon 2020 research and innovation program

under Zika- PLAN grant agreement No. 734584.

Ethics approval

Not applicable. This a review of published literature.

References

1. World Health Organization. Screening, assessment and management of neonates and infants with complications associated with Zika virus exposure in utero. Rapid Advice Guideline, 2016.

2. Ashwal S, Michelson D, Plawner L, Dobyns WB, Quality Standards Subcommittee of the American Academy of N, the Practice Committee of the Child Neurology S. Practice parameter: Evaluation of the child with microcephaly (an evidence-based review): report of the Quality Standards Subcommittee of the American Academy of Neurology and the Practice Committee of the Child Neurology Society. *Neurology* 2009; **73**(11): 887-97.

3. Leviton A, Holmes LB, Allred EN, Vargas J. Methodologic issues in epidemiologic studies of congenital microcephaly. *Early Hum Dev* 2002; **69**(1-2): 91-105.

4. Victora CG, Schuler-Faccini L, Matijasevich A, Ribeiro E, Pessoa A, Barros FC. Microcephaly in Brazil: how to interpret reported numbers? *Lancet* 2016; **387**(10019): 621-4.

5. World Health Organization. WHO Child Growth Standards.

http://www.who.int/childgrowth/en/.

6. Villar J, Cheikh Ismail L, Victora CG, et al. International standards for newborn weight, length, and head circumference by gestational age and sex: the Newborn Cross-Sectional Study of the INTERGROWTH-21st Project. *Lancet* 2014; **384**(9946): 857-68.

7. Morris JK, Rankin J, Garne E, et al. Prevalence of microcephaly in Europe: population based study. *BMJ* 2016; **354**: i4721.

8. Woods CG. Human microcephaly. *Curr Opin Neurobiol* 2004; **14**(1): 112-7.

9. Passemard S, Kaindl AM, Verloes A. Microcephaly. *Handb Clin Neurol* 2013; **111**: 129-41.

10. Barkovich AJ, Kuzniecky RI, Jackson GD, Guerrini R, Dobyns WB. A developmental and genetic classification for malformations of cortical development. *Neurology* 2005; **65**(12): 1873-87.

11. Szabo N, Pap C, Kobor J, Svekus A, Turi S, Sztriha L. Primary microcephaly in Hungary: epidemiology and clinical features. *Acta paediatrica* 2010; **99**(5): 690-3.

12. von der Hagen M, Pivarcsi M, Liebe J, et al. Diagnostic approach to microcephaly in childhood: a two-center study and review of the literature. *Dev Med Child Neurol* 2014; **56**(8): 732-41.

13. Graham KA, Fox DJ, Talati A, et al. Prevalence and Clinical Attributes of Congenital

Microcephaly - New York, 2013-2015. MMWR Morb Mortal Wkly Rep 2017; 66(5): 125-9.

14. Franca GV, Schuler-Faccini L, Oliveira WK, et al. Congenital Zika virus syndrome in Brazil: a case series of the first 1501 livebirths with complete investigation. *Lancet* 2016.

15. Oliveira Melo AS, Malinger G, Ximenes R, Szejnfeld PO, Alves Sampaio S, Bispo de Filippis AM. Zika virus intrauterine infection causes fetal brain abnormality and microcephaly: tip of the iceberg? *Ultrasound Obstet Gynecol* 2016; **47**(1): 6-7.

16. Dolk H. The predictive value of microcephaly during the first year of life for mental retardation at seven years. *Dev Med Child Neurol* 1991; **33**(11): 974-83.

17. Heinonen K, Raikkonen K, Pesonen AK, et al. Prenatal and postnatal growth and cognitive abilities at 56 months of age: a longitudinal study of infants born at term. *Pediatrics* 2008; **121**(5): e1325-33.

18. Woods CG, Parker A. Investigating microcephaly. Arch Dis Child 2013; 98(9): 707-13.

19. Krauss MJ, Morrissey AE, Winn HN, Amon E, Leet TL. Microcephaly: an epidemiologic analysis. *Am J Obstet Gynecol* 2003; **188**(6): 1484-9; discussion 9-90.

20. O'Rahilly R, Muller F. Developmental Stages in Human Embryos. Washington: Carnegie Institution of Washington Publication 637; 1987.

21. Chenn A, Walsh CA. Regulation of Cerebral Cortical Size by Control of Cell Cycle Exit in Neural Precursors. *Science* 2002; **297**(5580): 365-9.

22. Sahara S OLD. Fgf10 regulates transition period of cortical stem cell differentiation to radial glia controlling generation of neurons and basal progenitors. *Neuron* 2009; **63**: 48-62.

23. Buchman JJ TH, Zhou Y, Frank CL, Xie Z, Tsai LH. Cdk5rap2 interacts with pericentrin to maintain the neural progenitor pool in the developing neocortex. *Neuron* 2010; **66**: 386-402.

24. Marthiens V RM, Pennetier C, Tessier S, Paul-Gilloteaux P, Basto R. Centrosome amplification causes microcephaly. *Nat Cell Biol* 2013; **15**: 731-40.

Chen JF ZY, Wilde J, Hansen KC, Lai F, Niswander L. Microcephaly disease gene Wdr62 regulates mitotic progression of embryonic neural stem cells and brain size. *Nat Commun* 2014; 5: 3885.
 Lancaster MA RM, Martin CA, Wenzel D, Bicknell LS, Hurles ME, Homfray T, Penninger JM,

Jackson AP, Knoblich JA. Cerebral organoids model human brain development and microcephaly. . *Nature* 2013; **501**: 373-9.

27. Cooper LZ, Ziring PR, Ockerse AB, Fedun BA, Kiely B, Krugman S. Rubella-Clinical manifestations and management. *Am J Dis Child* 1969; **118**: 18-29.

28. Webster WS. Teratogen update: congenital rubella. *Teratology* 1998; **58**: 13-23.

29. Driggers RW, Ho CY, Korhonen EM, et al. Zika Virus Infection with Prolonged Maternal Viremia and Fetal Brain Abnormalities. *N Engl J Med* 2016; **374**(22): 2142-51.

30. Mlakar J, Korva M, Tul N, et al. Zika Virus Associated with Microcephaly. *N Engl J Med* 2016; **374**(10): 951-8.

31. Feldman B, Yinon Y, Tepperberg Oikawa M, Yoeli R, Schiff E, Lipitz S. Pregestational, periconceptional, and gestational primary maternal cytomegalovirus infection: prenatal diagnosis in 508 pregnancies. *American journal of obstetrics and gynecology* 2011; **4**: e1-6.

32. Lipitz S, Yinon Y, Malinger G, et al. Risk of cytomegalovirus-associated sequelae in relation to time of infection and findings on prenatal imaging. *Ultrasound Obstet Gynecol* 2013; **41**: 508-14.

33. McAuley JB. Congenital Toxoplasmosis J Pediatric Infect Dis Soc 2014; Suppl 1: S30–S5.

34. Marquez L, Levy ML, Munoz FM, Palazzi DL. A Report of Three Cases and Review of Intrauterine Herpes Simplex Virus Infection. *Pediatr Infect Dis J* 2011; **30**(2): 153-7.

35. Hutto C, Arvin A, Jacobs R, et al. Intrauterine herpes simplex virus infections. *J Pediatr* 1987; **110**(1): 97-101.

36. Barefoot KH, Little GA, Ornvold KT. Fetal Demise Due to Herpes Simplex Virus: An Illustrated Case Report. *Journal of Perinatology*

2002; 22: 86-8.

37. Odeberg J, Wolmer N, Falci S, Westgren M, Seiger A, Söderberg-Nauclér C. Human cytomegalovirus inhibits neuronal differentiation and induces apoptosis in human neural precursor cells. *J Virol* 2006; **80**: 8929-39.

38. Johansson AB, Rassart A, Blum D, Van Beers D, Liesnard C. Lower-limb hypoplasia due to intrauterine infection with herpes simplex virus type 2: possible confusion with intrauterine varicellazoster syndrome. *Clin Infect Dis* 2004; **38**(7): 57-62.

39. Lazar M, Perelygina L, Martines R, et al. Immunolocalization and Distribution of Rubella Antigen in Fatal Congenital Rubella Syndrome. *EBioMedicine* 2015; **3**: 86-92.

40. Dang J, Tiwari SK, Lichinchi G, et al. Zika Virus Depletes Neural Progenitors in Human Cerebral Organoids through Activation of the Innate Immune Receptor TLR3. *Cell Stem Cell* 2016; **19**(2): 258-65.

41. Qian X, Nguyen HN, Song MM, et al. Brain-Region-Specific Organoids Using Mini-bioreactors for Modeling ZIKV Exposure. *Cell* 2016; **165**(5): 1238-54.

42. Lafaille FG, Pessach IM, Zhang SY, et al. Impaired intrinsic immunity to HSV-1 in human iPSC-derived TLR3-deficient CNS cells. *Nature* 2012; **491**: 769-73.

43. Garcez PP, Loiola EC, Madeiro da Costa R, et al. Zika virus impairs growth in human neurospheres and brain organoids. *Science* 2016; **352**(6287): 816-8.

44. Luo MH, Hannemann H, Kulkarni AS, Schwartz PH, O'Dowd JM, Fortunato EA. Human cytomegalovirus infection causes premature and abnormal differentiation of human neural progenitor cells. *J Virol* 2010; **84**: 3528-41.

45. Li XJ, Liu XJ, Yang B, et al. Human Cytomegalovirus Infection Dysregulates the Localization and Stability of NICD1 and Jag1 in Neural Progenitor Cells. *J Virol* 2015; **89**: 6792-804.

46. D'Aiuto L, Di Maio R, Heath B, et al. Human induced pluripotent stem cell-derived models to investigate human cytomegalovirus infection in neural cells. *PLoS One* 2012; **7**: e49700.

47. Rolland M, Li X, Sellier Y, et al. PPARgamma Is Activated during Congenital Cytomegalovirus Infection and Inhibits Neuronogenesis from Human Neural Stem Cells. *PLoS Pathog* 2016; **12**(4): e1005547.

48. Braun E, Zimmerman T, Hur TB, et al. Neurotropism of herpes simplex virus type 1 in brain organ cultures. *J Gen Virol* 2006; **87**: 2827-37.

49. Hu S, Rotschafer JH, Lokensgard JR, Cheeran MC-J. Activated CD8+ T Lymphocytes Inhibit Neural Stem/Progenitor Cell Proliferation: Role of Interferon-Gamma. *PLoS ONE* 2014; 9(8): e105219.
50. Nguyen TV, Pham VH, Abe K. Pathogenesis of Congenital Rubella Virus Infection in Human Fetuses: Viral Infection in the Ciliary Body Could Play an Important Role in Cataractogenesis.

EBioMedicine 2014; **2**: 59-63.

51. Rorke LB, Fabiyi A, Elizan TS, Sever JL. Experimental cerebrovascular lesions in congenital and neonatal rubella-virus infections of ferrets. *Lancet* 1968; **2**: 153-4.

52. Tondury G, Smith DW. Fetal rubella pathology. *J Pediat* 1966; **68**: 867.

53. Cugola FR, Fernandes IR, Russo FB, et al. The Brazilian Zika virus strain causes birth defects in experimental models. *Nature* 2016; **534**: 267-71.

54. Tang H, Hammack C, Ogden SC, et al. Zika Virus Infects Human Cortical Neural Progenitors and Attenuates Their Growth. *Cell Stem Cell* 2016; (18): 587-90.

55. Hamel R, Liegeois F, Wichit S, et al. Zika virus: epidemiology, clinical features and host-virus interactions. *Microbes Infect* 2016; **18**(7-8): 441-9.

56. Nowakowski TJ, Pollen AA, Di Lullo E, andoval-Espinosa C, Bershteyn M, Kriegstein AR. Expression Analysis Highlights AXL as a Candidate Zika Virus Entry Receptor in Neural Stem Cells. *Cell Stem Cell* 2016; **5**: 591-6.

57. Onorati M, Li Z, Liu F, et al. Zika Virus Disrupts Phospho-TBK1 Localization and Mitosis in Human Neuroepithelial Stem Cells and Radial Glia. *Cell Rep* 2016; **16**(10): 2576-92.

58. Wells MF, Salick MR, Wiskow O, et al. Genetic Ablation of AXL Does Not Protect Human Neural Progenitor Cells and Cerebral Organoids from Zika Virus Infection. *Cell Stem Cell* 2016; **19**(6): 703-8.

59. Pillai S, Nguyen J, Johnson J, Haura E, Coppola D, Chellappan S. Tank binding kinase 1 is a centrosome-associated kinase necessary for microtubule dynamics and mitosis. *Nat Commun* 2015; **6**: 10072.

60. Li C, Xu D, Ye Q, et al. Zika Virus Disrupts Neural Progenitor Development and Leads to Microcephaly in Mice. *Cell Stem Cell* 2016; **19**: 120-6.

61. Wu KY, Zuo GL, Li XF, et al. Vertical transmission of Zika virus targeting the radial glial cells affects cortex development of offspring mice. *Cell Res* 2016; **26**: 645-54.

62. Adams Waldorf KM, Stencel-Baerenwald JE, Kapur RP, et al. Fetal brain lesions after subcutaneous inoculation of Zika virus in a pregnant nonhuman

primate. Nat Med 2016; 22(11): 1256-9.

63. Manicklal S, Emery VC, Lazzarotto T, Boppana SB, Gupta RK. The "silent" global burden of congenital cytomegalovirus. *Clinical microbiology reviews* 2013; **26**(1): 86-102.

64. Sheevani, Jindal N, Aggarwal A. A pilot seroepidemiological study of cytomegalovirus infection in women of child bearing age. *Indian journal of medical microbiology* 2005; **23**(1): 34.

65. Schoenfisch AL, Dollard SC, Amin M, et al. Cytomegalovirus (CMV) shedding is highly correlated with markers of immunosuppression in CMV-seropositive women. *Journal of medical microbiology* 2011; **60**(6): 768-74.

66. Cannon MJ, Schmid DS, Hyde TB. Review of cytomegalovirus seroprevalence and demographic characteristics associated with infection. *Reviews in medical virology* 2010; **20**(4): 202-13.

67. Alanen A, Kahala K, Vahlberg T, Koskela P, Vainionpää R. Seroprevalence, incidence of prenatal infections and reliability of maternal history of varicella zoster virus, cytomegalovirus, herpes simplex virus and parvovirus B19 infection in South-Western Finland. *BJOG: An International Journal of Obstetrics & Gynaecology* 2005; **112**(1): 50-6.

68. Seo S, Cho Y, Park J. Serologic screening of pregnant Korean women for primary human cytomegalovirus infection using IgG avidity test. *Korean J Lab Med* 2009; **29**(6): 557-62.

69. Kenneson A, Cannon MJ. Review and meta-analysis of the epidemiology of congenital cytomegalovirus (CMV) infection. *Reviews in medical virology* 2007; **17**(4): 253-76.

70. Ornoy A, Diav-Citrin O. Fetal effects of primary and secondary cytomegalovirus infection in pregnancy. *Reproductive toxicology* 2006; **21**(4): 399-409.

71. Picone O, Vauloup-Fellous C, Cordier AG, et al. A series of 238 cytomegalovirus primary infections during pregnancy: description and outcome. *Prenatal diagnosis* 2013; **33**(8): 751-8.

72. Britt W. Controversies in the natural history of congenital human cytomegalovirus infection: the paradox of infection and disease in offspring of women with immunity prior to pregnancy. *Med Microbiol Immunol* 2015; **204**(3): 263-71.

73. Society for Maternal-Fetal M, Hughes BL, Gyamfi-Bannerman C. Diagnosis and antenatal management of congenital cytomegalovirus infection. *American journal of obstetrics and gynecology* 2016; **214**(6): B5-B11.

74. Khamduang W, Jourdain G, Sirirungsi W, et al. The interrelated transmission of HIV-1 and cytomegalovirus during gestation and delivery in the offspring of HIV-infected mothers. *Journal of acquired immune deficiency syndromes* 2011; **58**(2): 188-92.

75. Pinninti SG, Kimberlin DW. Neonatal herpes simplex virus infections. *Pediatr Clin North Am* 2013; **60**(2): 351-65.

76. Looker KJ, Garnett GP, Schmid GP. An estimate of the global prevalence and incidence of herpes simplex virus type 2 infection. *Bulletin of the World Health Organization* 2008; **86**(10): 805-12A.

77. Looker KJ, Magaret AS, Turner KM, Vickerman P, Gottlieb SL, Newman LM. Global estimates of prevalent and incident herpes simplex virus type 2 infections in 2012. *PLoS One* 2015; **10**(1): e114989.

78. Vynnycky E, Adams EJ, Cutts FT, et al. Using Seroprevalence and Immunisation Coverage Data to Estimate the Global Burden of Congenital Rubella Syndrome, 1996-2010: A Systematic Review. *PloS one* 2016; **11**(3): e0149160.

79. World Health Organization. WHO | Rubella fact sheet. *WHO* 2016.

80. Pandolfi E, Gesualdo F, Rizzo C, et al. Global seroprevalence of rubella among pregnant and childbearing age women: a meta-analysis. *Eur J Public Health* 2017.

81. Tenter AM, Heckeroth AR, Weiss LM. Toxoplasma gondii: from animals to humans. *Int J Parasitol* 2000; **30**(12-13): 1217-58.

82. Jones JL, Kruszon-Moran D, Wilson M, McQuillan G, Navin T, McAuley JB. Toxoplasma gondii infection in the United States: seroprevalence and risk factors. *American journal of epidemiology* 2001; **154**(4): 357-65.

83. Torgerson PR, Mastroiacovo P. The global burden of congenital toxoplasmosis: a systematic review. *Bulletin of the World Health Organization* 2013; **91**(7): 501-8.

84. Wang Z-D, Wang S-C, Liu H-H, et al. Prevalence and burden of Toxoplasma gondii infection in HIV-infected people: a systematic review and meta-analysis. *The Lancet HIV*; **4**(4): e177-e88.

85. Duffy MR, Chen T-H, Hancock WT, et al. Zika Virus Outbreak on Yap Island, Federated States of Micronesia. *New England Journal of Medicine* 2009; **360**(24): 2536-43.

86. Aubry M, Teissier A, Huart M, et al. Zika virus seroprevalence, French Polynesia, 2014–2015. *Emerging infectious diseases* 2017.

87. World Health Organization. WHO | Zika situation report. WHO 2017.

88. Coelho FC, Durovni B, Saraceni V, et al. Sexual transmission causes a marked increase in the incidence of Zika in women in Rio de Janeiro, Brazil. *bioRxiv* 2016: 055459.

89. Health Protection Agency Rash Guidance Working Group. Guidance on Viral Rash in Pregnancy. Investigation, Diagnosis and Management of Viral Rash Illness, or Exposure to Viral Rash Illness, in Pregnancy. London: Public Health England, 2011.

90. Pass RF, Boppana S, Jeffries DJ, Hudson CN. Viral infection in obstetrics and gynaecology. New York: Arnold; 1999.

91. Best JM. Rubella. Semin Fetal Neonatal Med 2007; 12(3): 182-92.

92. Mendelson E, Aboudy Y, Smetana Z, Tepperberg M, Grossman Z. Laboratory assessment and diagnosis of congenital viral infections: Rubella, cytomegalovirus (CMV), varicella-zoster virus (VZV), herpes simplex virus (HSV), parvovirus B19 and human immunodeficiency virus (HIV). *Reproductive toxicology* 2006; **21**(4): 350-82.

93. Lazzarotto T, Guerra B, Lanari M, Gabrielli L, Landini MP. New advances in the diagnosis of congenital cytomegalovirus infection. *J Clin Virol* 2008; **41**(3): 192-7.

94. DPDx Laboratory Identification of Parasitic Diseases of Public Health Concern. Toxoplasmosis. 2015. <u>http://www.cdc.gov/dpdx/toxoplasmosis/dx.html</u> (accessed 26/10/16).

95. Petersen E. Toxoplasmosis. Semin Fetal Neonatal Med 2007; 12(3): 214-23.

96. Dard C, Fricker-Hidalgo H, Brenier-Pinchart MP, Pelloux H. Relevance of and New

Developments in Serology for Toxoplasmosis. Trends in parasitology 2016; 32(6): 492-506.

97. Rorman E, Zamir CS, Rilkis I, Ben-David H. Congenital toxoplasmosis--prenatal aspects of Toxoplasma gondii infection. *Reproductive toxicology* 2006; **21**(4): 458-72.

98. Alanen A, Hukkanen V. Herpes simplex virus DNA in amniotic fluid without neonatal infection. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* 2000; **30**(2): 363-7.

99. Calvet G, Aguiar RS, Melo AS, et al. Detection and sequencing of Zika virus from amniotic fluid of fetuses with microcephaly in Brazil: a case study. *Lancet Infect Dis* 2016; **16**(6): 653-60.

100. World Health Organization. Immunization coverage, 2016.

101. McLeod R, Kieffer F, Sautter M, Hosten T, Pelloux H. Why prevent, diagnose and treat congenital toxoplasmosis? *Mem Inst Oswaldo Cruz* 2009; **104**(2): 320-44.

102. Cheeran MC, Lokensgard JR, Schleiss MR. Neuropathogenesis of congenital cytomegalovirus infection: disease mechanisms and prospects for intervention. *Clinical microbiology reviews* 2009; **22**(1): 99-126, Table of Contents.

103. Dollard SC, Grosse SD, Ross DS. New estimates of the prevalence of neurological and sensory sequelae and mortality associated with congenital cytomegalovirus infection. *Reviews in medical virology* 2007; **17**(5): 355-63.

104. Dreher AM, Arora N, Fowler KB, et al. Spectrum of disease and outcome in children with symptomatic congenital cytomegalovirus infection. *The Journal of pediatrics* 2014; **164**(4): 855-9.

105. Kimberlin DW. Neonatal Herpes Simplex Infection. *Clinical microbiology reviews* 2004; 17(1):1-13.

106. Miller E, Cradock-Watson JE, Pollock TM. Consequences of confirmed maternal rubella at successive stages of pregnancy. *The Lancet* 1982; **8302**: 781-4.

107. Givens KT, Lee DA, Jones T, Ilstrup DM. Congenital rubella syndrome: ophthalmic manifestations and associated systemic disorders. *Br J Ophthalmol* 1993; **77**(6): 358-63.

108. Reef SE, Plotkin S, Cordero JF, et al. Preparing for Elimination of Congenital Rubella Syndrome (CRS): Summary of a Workshop on CRS Elimination in the United States. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* 2000; **31**(1): 85-95.

109. Syrocot study group, Thiebaut R, Leproust S, Chene G, Gilbert R. Effectiveness of prenatal treatment for congenital toxoplasmosis: a meta-analysis of individual patients' data. *Lancet* 2007; **369**(9556): 115-22.

110. Cortina-Borja M, Tan HK, Wallon M, et al. Prenatal treatment for serious neurological sequelae of congenital toxoplasmosis: an observational prospective cohort study. *PLoS Med* 2010; **7**(10).

111. Capobiango JD, Bregano RM, Navarro IT, et al. Congenital toxoplasmosis in a reference center of Parana, Southern Brazil. *Braz J Infect Dis* 2014; **18**(4): 364-71.

112. Cauchemez S, Besnard M, Bompard P, et al. Association between Zika virus and microcephaly in French Polynesia, 2013–15: a retrospective study. *The Lancet* 2016; **387**(10033): 2125-32.

113. Brasil P, Pereira JP, Jr., Raja Gabaglia Č, et al. Zika Virus Infection in Pregnant Women in Rio de Janeiro - Preliminary Report. *N Engl J Med* 2016.

114. Moore CA, Staples JE, Dobyns WB, et al. Characterizing the Pattern of Anomalies in Congenital Zika Syndrome for Pediatric Clinicians. *JAMA Pediatr* 2017; **171**(3): 288-95.

115. Melo AS, Aguiar RS, Amorim MM, et al. Congenital Zika Virus Infection: Beyond Neonatal Microcephaly. *JAMA Neurol* 2016; **73**(12): 1407-16.

116. Ventura CV, Maia M, Dias N, Ventura LO, Belfort R, Jr. Zika: neurological and ocular findings in infant without microcephaly. *Lancet* 2016; **387**(10037): 2502.

117. Reynolds MR, Jones AM, Petersen EE, et al. Vital Signs: Update on Zika Virus-Associated Birth Defects and Evaluation of All U.S. Infants with Congenital Zika Virus Exposure - U.S. Zika Pregnancy Registry, 2016. *MMWR Morb Mortal Wkly Rep* 2017; **66**(13): 366-73.

118. Noyola DE, Demmler GJ, Nelson CT, et al. Early predictors of neurodevelopmental outcome in symptomatic congenital cytomegalovirus infection. *The Journal of pediatrics* 2001; **138**(3): 325-31.

119. Abdel Razek AA, Kandell AY, Elsorogy LG, Elmongy A, Basett AA. Disorders of cortical formation: MR imaging features. *AJNR Am J Neuroradiol* 2009; **30**(1): 4-11.

120. World Health Organization. Psychosocial support for pregnant women and for families with microcephaly and other neurological complications in the context of Zika virus. Interim guidance for health-care providers, 2016.

121. Centers for Disease Control. Facts about Microcephaly. 2016.

http://www.cdc.gov/ncbddd/birthdefects/microcephaly.html (accessed 15 July 2016).

122. Holt RL, Mikati MA. Care for child development: basic science rationale and effects of interventions. *Pediatr Neurol* 2011; **44**(4): 239-53.

123. Britto PR, Lye SJ, Proulx K, et al. Nurturing care: promoting early childhood development. *Lancet* 2016.

124. UNICEF. Violence against Children with Disabilities. Report for UNICEF/United Nations Secretary General's Office. New York: UNICEF, 2005.

125. World Health Organization, World Bank. World Report on Disability, 2011.

126. UNICEF. Children with Disabilities, 2012.

127. Groce N. Disability, Public Health and Social Injustice. In: Levy B, Sidel, V., ed. Social Injustice and Public Health. 2nd ed. Oxford: Oxford University Press; 2014.

128. Yamin A. Health, Human Rights and the Zika Virus. *Health and Human Rights* 2016.

129. Palmer M. Disability and Poverty: A Conceptual Review. *Journal of Disability Policy Studies* 2011; **21**(4): 210-8.

130. Mitra S, Posarac A, Vick B. Disability and Poverty in Developing Countries: A Multidimensional Study. *World Development* 2013; **41**: 1-18.

131. United Nations. UN Convention on the Rights of Persons with Disabilities. 2006.

132. Klein JO, Remington JS. Current concepts of infections of the fetus and newborn infant In: Remington JS, Klein JO, eds. Infectious Disease of the Fetus and Newborn Infant. 4th ed. Philadelphia: WB Saunders Co; 1995: 1–16.

Johnson KE. Overview of TORCH infections. In: Weisman LE, Edwards MS, eds. UptoDate;
 2015.

134. Leruez-Ville M, Ville Y. Optimum treatment of congenital cytomegalovirus infection. *Expert Rev Anti Infect Ther* 2016; **14**(5): 479-88.

135. Kimberlin DW, Baley J, Committee on infectious d, Committee on f, newborn. Guidance on management of asymptomatic neonates born to women with active genital herpes lesions. *Pediatrics* 2013; **131**(2): e635-46.

136. Lee J, Bowden DS. Rubella Virus Replication and Links to Teratogenicity *Clinical microbiology reviews* 2000; **13**(4): 571–87.

137. Silasi M, Cardenas I, Kwon J-Y, Racicot K, Aldo P, Mor G. Viral infections during pregnancy. *American Journal of Reproductive Immunology* 2015; **73**(3): 199-213.

138. Montoya JG, Liesenfeld O. Toxoplasmosis. The Lancet 2004; 363(9425): 1965-76.

139. Wikan N, Smith DR. Zika virus: history of a newly emerging arbovirus. *Lancet Infect Dis* 2016; **16**(7): e119-e26.

140. Waddell LA, Greig JD. Scoping Review of the Zika Virus Literature. *PLoS One* 2016; **11**(5): e0156376.

141. Bharucha T, Breuer J. Review: A neglected Flavivirus: an update on Zika virus in 2016 and the future direction of research. *Neuropathol Appl Neurobiol* 2016; **42**(4): 317-25.

142. Petersen LR, Jamieson DJ, Powers AM, Honein MA. Zika Virus. *N Engl J Med* 2016; **374**(16): 1552-63.

143. Kimberlin DW, Jester PM, Sanchez PJ, et al. Valganciclovir for symptomatic congenital cytomegalovirus disease. *N Engl J Med* 2015; **372**(10): 933-43.

144. Kimberlin DW, Lin CY, Sanchez PJ, et al. Effect of ganciclovir therapy on hearing in symptomatic congenital cytomegalovirus disease involving the central nervous system: a randomized, controlled trial. *The Journal of pediatrics* 2003; **143**(1): 16-25.

145. Shah T, Luck S, Sharland M, Kadambari S, Heath P, Lyall H. Fifteen-minute consultation: diagnosis and management of congenital CMV. *Archives of disease in childhood Education and practice edition* 2016.

146. Kimberlin DW, Lin CY, Jacobs RF, et al. Safety and efficacy of high-dose intravenous acyclovir in the management of neonatal herpes simplex virus infections. *Pediatrics* 2001; **108**(2): 230-8.

147. Plotkin SA, Reef SE, Cooper LZ, Alford CA. Rubella. In: Remington JS, Klein JO, Wilson CB, Baker CJ, eds. Infectious Diseases of the Fetus and Newborn Infant. 7th ed. Philadelphia: Elsevier Saunders; 2011: 861.

148. Daffos F, Forestier F, Capella-Pavlovsky M, et al. Prenatal management of 746 pregnancies at risk for congenital toxoplasmosis. *N Engl J Med* 1988; **318**(5): 271-5.

149. R Development Core Team. R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing. Vienna, Austria 2011.